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Anti-inflammatory properties of Rosmarinic acid - A review

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ABSTRACT

Phytochemicals are natural, non nutritive chemicals rich in pharmacological activities. They show a very potent antioxidant activity and this activity is closely linked to various beneficial actions, including anti-aging, prevention of cancer, cardiovascular disease, etc. Medicinal plants play a vital role in drug discovery that can be used to cure various ailments in humans. A safe new phytochemical compound as drugs is one of the major searches world-wide. Rosmarinic acid, a polyphenolic compound is generally available in the wide species of the Boraginaceae and Lamiaceae family (Nepetoideae subfamily). The phytochemical has two phenolic rings with ortho positioned hydroxyl groups. A carboxylic acid, carbonyl group and an unsaturated double bond is present between the two phenolic rings. The compound is thus an ester linked caffeic acid and 3, 4-dihydroxyphenyllactic acid. It has a broad range of biological activity that includes antiviral, antibacterial, antitumor, antihepatitis, anti-inflammation and inhibiting blood clots. Inflammation, a localized protective response elicited by injury or destructions of tissues, is mediated by various proinflammatory cytokines, is associated with every health condition. Inflammation is also important cause for many allergic diseases. Rosmarinic acid, being one of the promising active principles, can be developed for various pharmacological activities through clinical trials and applications further. This review aimed to describe the antiinflammatory properties of rosmarinic acid in various diseased conditions and to understand its basic mechanisms.

Keywords: Anti-inflammatory properties; Phytochemicals; inflammation; Rosmarinic acid.

INTRODUCTION

The mammalian tissues show inflammatory response against harmful stimuli caused by hostile agents such as toxic chemical substances, physical injury, infectious organisms, tumor growth etc, (Sobota et al., 2000) further local accumulation of plasmic fluid and blood cells occurs. Inflammation can be either acute or chronic conditions. The process of Inflammation is characterized by the production of mediators such as histamine, bradykinin, prostaglandins, platelet-activating factor, leukotrienes and the release of chemicals from tissues and migrating cells (Tomlinson et al., 1994; Cuzzocrea et al., 2004). Prostanoids is synthesized by Cyclooxygenase (COX), it is involved in platelet aggregation, pain and inflammation (Pilotto et al., 2010). Central role of inflammatory responses is through p38a MAPK, a serine / threonine kinase. It is involved in the biosynthesis and release of pro-inflammatory cytokines e.g. IL-1 β and TNF α . The production is controlled at the level of transcription and the translation (Laufer et al.,

* Corresponding Author Email: drsridevimuruhan@gmail.com Contact: +91-9942364284, +91-9843526815 Received on: 27.06.2017 Revised on: 11.11.2017 Accepted on: 14.11.2017 2002). These reaction mediators provoke, maintain and aggravate many disorders. The key feature of inflammatory activity is production of reactive species by phagocytic cells that injures cell and tissue by oxidative degradation of essential cellular components (Halliwell et al., 1988).



Figure 1: Rosmarinic acid - Chemical Structure

Plants are potential sources for producing new drugs to treat many chronic and infectious diseases. The parts of many medicinal plants possess bioactive compounds that have varied medicinal properties; hence they are used as raw drugs (Mahesh et al., 2008). Phytochemicals are low molecular weight active secondary metabolites in plants, few examples are caffeine, nicotine, ginkgo, ginseng, valerian, curcumin, resveratrol, epigallocatechin-3-gallate etc. (Kennedy et al., 2011). They are the potential sources of natural anti-inflammatory, antioxidant, antimicrobial, anticancer, hepatoprotective and nephroprotective activities (Parnham et al., 1985; Petersen M., 1994). They protect the plant against ultraviolet radiation, pathogens, and herbivores (Harborne et al., 2000). Also they are used to effectively treat the various ailments for mankind. In recent years, the importance of medicinal plants for treating many diseased conditions were increasing. They exhibit potent pharmacological properties that are proved by different epidemiological and in vitro/in vivo experimental studies. The phytochemicals such as epigallocatechin gallate, resveratrol, phytosterol, myricetin, gingerol, etc., directly impacts various molecular signal transduction pathways like oxidative stress, cell proliferation/migration, inflammation cascade and metabolic disorders that are involved in the development of diseased conditions (Upadhyay et al., 2015).

Rosmarinic acid

In Boraginaceae species and in the Nepetoideae subfamily of the Lamiaceae herbs, (rosemary, sweet basil, perilla) a polyphenolic compound, Rosmarinic acid (RA) is found (del Bano et al., 2003; Kintzios et al., 2003; Osakabe et al., 2002) with biological activities. RA is also found in other plant kingdoms which taxonomically belong to non-related families (Holzmannova 1996; Petersen 2003). It (α-o-caffeoyl-3,4-dihydroxyphenyl lactic acid; RA), occurs in the form of hydroxylated phenolic compound naturally as an ester of caffeic acid and 3, 4- dihydroxy phenyl lactic acid. RA posses extensive pharmacological activities such as anti-oxidant, anti- inflammatory, anti-tumor and anti-angiogenic activities (Osakabe et al., 2004; McKay et al., 2006; Huang et al., 2006; Kosar et al., 2008). In addition they causes attenuation of T cell receptor mediated signaling, blocking of complement fixation and suppression of inhibitor of nuclear factor kappa-B (IKK-β) downstream signaling during the upregulation of C-C motif chemokine 11(CCL11) induced by tumor necrosis factor (TNF)- α , CCL11 is a potent chemo attractant and an activator of T helper 2 cells, eosinophils and basophils (Sahu et al., 1999; Lee et al., 2006; Kang et al., 2003). Approximately 3% RA was found in medicinal plants by dry weight (Lamaison et al., 1989).

The anti-inflammatory properties of rosmarinic acid is due to the inhibition of activity of enzymes such as lipoxygenase (LOX) and cyclooxygenases (COX), inhibition of inflammatory cytokines expression (Sanbongi et al., 2003) and alteration in the complement cascade pathway (Parnham et al., 1985; Mirzoeva et al., 1996; Krol et al., 1996; Petersen et al, 2003). Basic mechanism underlying in anti-inflammatory and antiatheroesclerotic activity of rosmarinic acid by *in-vitro* and *in-vivo* models confirm high specific antioxidant activity with less production of pro-inflammatory molecules and also by preventing the low density lipoprotein oxidation. This review focuses on the reported antiinflammatory activities of rosmarinic acid.

Reported anti-inflammatory activities of Rosmarinic acid

Lung injury

Exposure of animals as well as humans to diesel exhaust particles (DEP) induces an acute inflammatory response (Salvi et al., 1999), by generating reactive oxygen species (Sagai et al., 1993 & 1996; Ichinose et al., 1995; Lim et al., 1998), by inducing the production of P450 reductase and/or P450 in the inflammatory process (Kumagai et al., 1997; Lim et al., 1998; Takano et al., 2002). Sanbongi et al., (2003) observed that exposure of male ICR mice to DEP caused pathophysiological changes such as neutrophilic inflammation and edema in the lung by decreasing the local expression of keratinocyte chemoattractant, interleukin -1ß, monocyte chemoattractant protein -1 and macrophage inflammatory protein -1α . Rosmarinic acid is found to inhibit the expression of iNOS mRNA and formation of nitrotyrosine, 8-hydroxyguanosine that is enhanced by exposure to DEP. These effects also reduce the expression of proinflammatory cytokine and chemokines, which play a major role in inflammatory response in initiation and progression. The inflammatory response to allergens in the lungs of allergic asthma is a conseguence of increased expression of several inflammatory proteins like interleukin-4 in lung tissue and it is also associated with infiltration of the airway wall by eosinophils (inflammatory cells) (Lee et al., 2010).

Kang et al., (2003) reported that *Ocimum gratissimum* has rich contents of polyphenols, such as rosmarinic acid (Ola et al., 2009) and possess immunomodulatory activity by suppressing T-cell receptor (TCR) signaling. 100 mg/Kg methanolic extract of *Ocimum gratissimum* and 200 mg/Kg RA was analyzed by Costa et al., (2012) for the treatment of murine model of respiratory allergy. There was reduction in the leukocytes/eosinophils numbers, eosinophil peroxidase activity in bronchoalveolar lavage (BAL), presence of mucus in respiratory tract, histopathological changes in the lung, and interleukin - 4 in BAL suggesting their therapeutic potential.

Corsini et al., (2005) identified that pleurisy induced by intrapleural administration of carrageenan leads to immediate recruitment of polymorphonuclear cells (PMN) in the pleural space. The respiratory pathway inflammation is also due to mediators such as cytokines, histamine, leukotrienes, nitric oxide release and thromboxane A2 (Eum et al., 2003). Gamaro etal., (2011) proved that migration of polymorphonuclear and mononuclear cells stimulated by carrageenan into pleural space was reduced by caffeic acid and rosmarinic acid, suggesting that the anti-inflammatory property of rosmarinic acid in carrageenan induced rats is due to its breakdown products, like caffeic acid.

Hepatoprotective activity

The hepatoprotective effects of rosmarinic acid have been proved on liver ischemia/reperfusion injury in rats. The treatment with rosmarinic acid reduced neutrophil infiltration, hepatocellular damage and oxidative/nitrosative stress parameters caused by the injury. It also attenuated nuclear factor - κ B activation and down-regulated interleukin-1 β and tumor necrosis factor - α gene expression, the liver content of eNOS/iNOS and NO was also decreased. These data by Ramalho et al., (2014) indicates that in the ischemic liver, rosmarinic acid protects hepatocytes against ischemia/reperfusion injury by its anti-inflammatory and antioxidant effects.

The anti-inflammatory properties of rosmarinic acid from Rosmarinus officinalis in local inflammation of paw oedema rat model induced by carrageenin was assessed by Rocha et al, (2015). They found that by 6 hr by 60% of paw oedema has been reduced at the dose of 25 mg/kg of rosmarinic acid (Boonyarikpunchai et al., 2014). They also evaluated that, in systemic inflammation of liver ischaemia-reperfusion of rat models, there was a remarkable reduction in the serum concentration of transaminases and lactate dehydrogenase due to the prior administration of rosmarinic acid. Rosmarinic acid also reduced multi-organ dysfunction markers in liver, kidney and lung by modulating metalloproteinase-9 and nuclear factor- kB in the thermal injury rat model. In vivo studies also shown that rosmarinic acid will inhibits cobra venom factor induced paw edema (Leyck et al, 1983; Englberger et al, 1988).

Nephroprotective activities

Nephrotoxicity is the most serious dose-limiting side effects of chemotherapy with Cisplatin (CP) for cancer in bladder, testicular, ovarian, cervical, esophageal, head and neck (Giaccone 2000). The oxidative stress, tubular necrosis, inflammation and apoptosis are the major mechanisms underlying CP-induced nephrotoxicity (Miller et al., 2010). Domitrovic et al., (2014) examined that the treatment of nephrotoxicity induced by cisplatin (CP) in mice models with rosmarinic acid showed antiapoptotic activity by reducing p53, expression of active caspase-3 and phosphorylated p53. CP also cause considerable histopathological changes, increased blood urea nitrogen and serum creatinine. RA inhibited the tumor necrosis factor- α and nuclear factor-kappa B expression, and reduced the oxidative stress induced by CP, indicating the rosmarinic acid anti-inflammatory effect.

Pro-inflammatory mediators Inhibition

Pro-inflammatory mediators such as Tumor Necrosis Factor $-\alpha$, Interleukin-1 β & 6 possess many biological activities associated to chronic inflammatory diseases (Okada et al., 1998). Through the arachidonic acid cascade catalysed by cyclooxygenase isoforms, COX-1 and COX-2, inflammatory mediator Prostaglandin E2 (PGE2) was produced. Intracellular glutathione depletion, lipid peroxidation as well as production of reactive oxygen species in Lipopolysaccharide (LPS) induced human gingival fibroblasts cells was reduced by *Prunella vulgaris* L. extract and rosmarinic acid. They also suppressed, expression of inducible nitric oxide synthase and inhibited LPS-induced up-regulation of interleukin - 1β , interleukin – 6 and tumor necrosis factor- α .

Zdarilova et al., (2009) showed LPS-induced inflammatory process in gingival fibroblasts, including periodontal disease can be suppressed and modulated by *Prunella vulgaris* L. extract and rosmarinic acid. LPS mimicks an inflammatory response *in vitro* by stimulating macrophage cells to produce PGE2 (Patel et al., 1999). Meehye (2012) reported that rosmarinic acid is the key phytochemical in ethanol extracts of *Prunella vulgaris* which inhibits the production of PGE2 in RAW264.7 mouse cell line showing the anti-inflammatory properties.

Inhibition of diabetes-induced damage

Chronic hyperglycemia accelerates the advanced glycation end products (AGEs) formation and accumulation in various tissues. Diabetic vascular complications study showed that inflammatory state triggered by hyperglycaemia is by means of AGEs and their receptors together with Toll-like receptors and high mobilitygroup box-1 protein (Nogueira-Machado et al., 2011). mRNA of proinflammatory mediators like tumour necrosis factor- α and interleukin-1 β , was increased in aorta of diabetic rats induced by streptozocin (Sotnikova et al., 2013). Rosmarinic acid protects aortas of endothelial injury induced by diabetes by controlling the overexpression of proinflammatory mediators.

Seasonal Allergic Rhinoconjunctivitis (SAR) Inhibition by Rosmarinic acid

Rosmarinic acids inhibit Polymorphonuclear Leukocyte Infiltration and have effective control of seasonal allergic rhinoconjunctivitis. Takano et al., (2004) determined that in nasal lavage fluid of seasonal allergic rhinoconjunctivitis patients, there was significant reduction in the production of eosinophils and neutrophils, after oral supplementation with rosmarinic acid obtained from *Perilla frutescens* and inhibited PMNL infiltration into the nostrils, thus exhibiting the antiinflammatory properties.

Inhibition of epidermal inflammatory responses

Osakabe et al., (2004) found that topical application of the extract of perilla containing 68% rosmarinic acid or commercially available rosmarinic acid with equivalent quantity, showed anti-inflammatory activity in twostage skin carcinogenesis model (initiated by application of 7,12-dimethylbenz[a]anthracene and promoted by application of 12-tetradecanoylphorbol 13-acetate). RA treatment showed noticeable inhibition of neutrophil infiltration, decreased activity of myeloperoxidase, a neutrophil recruitment marker, reduced vascular cell adhesion molecule-1 and intercellular adhesion molecule 1 mRNA expression levels. Decrease in the levels of synthesis of chemokine, macrophage inflammatory protein-2, leukotriene B4 and Prostaglandin E2 were also observed with perilla extract.

Anti-inflammatory effects on Japanese Encephalitis

In children, acute encephalopathy is caused by Japanese encephalitis virus (JEV) that particularly targets the central nervous system (Chambers et al., 1997). In serum and cerebrospinal fluid, JEV infection showed elevated levels of inflammatory mediators such as Interleukins - 6 & 8, tumor necrosis factor alpha (TNF)-α and RANTES (Regulated on Activation, Normal T Cell Expressed and Secreted) (Ravi, et al., 1997; Kolson, et al., 1998; Singh, et al., 2000). JEV infection also sways the viral pathogenesis by stimulating microglial activation and proinflammatory mediators (IL-6, Monocyte Chemo attractant Protein-1 and TNF- α) (Ghoshal et al., 2007). Swarup et al., (2007) observed that rosmarinic acid significantly reduced the mortality of JEV infected mice and also decreased the viral loads and proinflammatory cytokine levels.

RA Inhibits complement factors

The classical and alternative pathways of complement activation were inhibited by rosmarinic acid (Peake et al, 1991; Englberger et al., 1988), it also stimulate prostacyclin synthesis via complement factors (Rampart et al., 1986) thereby proving its anti-inflammatory property.

CONCLUSION

The inflammation process is a complex network involved by different proinflammatory mediators. Oxidative stress and neutrophil oxidative burst initiates and mediates the inflammatory process and tissue injury. RA has been found to be effective against a number of diseases such as lung injury, liver injury, kidney disorders and other inflammatory disorders. The inhibition of the proinflammatory mediator's synthesis by lipoxygenase and cyclooxygenases enzymes activity, inhibition and prevention of reactive oxygen metabolites that promotes direct toxicity to activate the inflammation process are the major causes for antiinflammatory effect of the rosmarinic acid. The effect of rosmarinic acid in in-vitro and in-vivo models shows that the compound may be useful in the treatment of inflammation and oxidative stress. A deeper insight on the pharmacological activity and its mechanism with promising results is required to show that rosmarinic acid is a potential therapeutic compound for treating various inflammatory diseases.

REFERENCES

Boonyarikpunchai, W., Sukrong, S., Towiwat, P. Antinociceptive and anti-inflammatory effects of rosmarinic acid isolated from *Thunbergia laurifolia* Lindl. Pharmacol Biochem Behav. vol.124, 2014 pp. 67-73.

- Chambers, TJ., Tsai, TF., Pervikov, Y., Monath, TP. Vaccine development against dengue and Japanese encephalitis: report of a World Health Organization meeting. Vaccine. vol. 15, no. 14, 1997 pp.1494-1502.
- Corsini, E., Di Paola, R., Viviani, B., Genovese, T., Mazzon, E., Lucchi, L., Marinovich, M., Galli, C.L., Cuzzocrea, S. Increased carrageenan-induced acute lung inflammation in old rats. Immunology. vol. 115, no. 2, 2005 pp.253-261.
- Costa, RS., Carneiro, TCB., Cerqueira-Lima, AT., Queiroz, NV., Alcântara-Neves, NM., Pontes-de-Carvalho, LC., da Silva Velozo, E., Oliveira, EJ., Figueiredo, CA. *Ocimum gratissimum* Linn. and rosmarinic acid, attenuate eosinophilic airway inflammation in an experimental model of respiratory allergy to Blomia tropicalis. International immunopharmacology. vol.13, no.1, 2012 pp.126-134.
- Cuzzocrea, S., Pisano, B., Dugo, L., Lanaro, A., Maffia, P., et al., "Rosiglitazone, a ligand of the peroxisome proliferator-activated receptor- γ 3, reduces acute inflammation," *European Journal of Pharmacology*, vol. 483, no. 1, 2004 pp. 79–93.
- del Bano, MJ., Lorente, J., Castillo, J., Benavente-García, O., del Rio, JA., Ortuño, A., Quirin, KW., Gerard, D. Phenolic diterpenes, flavones, and rosmarinic acid distribution during the development of leaves, flowers, stems, and roots of *Rosmarinus officinalis*. Antioxidant activity. J Agric Food Chem, vol. 51, no.15, 2003 pp. 4247-4253.
- Domitrovic, R., Potocnjak, I., Crncevic-Orlic, Z., Skoda, M. Nephroprotective activities of rosmarinic acid against cisplatin-induced kidney injury in mice. Food and Chemical Toxicology. vol. 66, 2014 pp. 321-328.
- Englberger, W., Hadding, U., Etschenberg, E., Graf, E., Leyck, S., Winkelmann, J., Parnham, MJ. Rosmarinic acid: a new inhibitor of complement C3-convertase with anti-inflammatory activity. International journal of immunopharmacology. vol.10, no.6, 1988 pp. 729-737.
- Eum, SY., Maghni, K., Hamid, Q., Eidelman, DH., Campbell, H., Isogai, S. Martin, JG. Inhibition of allergic airways inflammation and airway hyperresponsiveness in mice by dexamethasone: role of eosinophils, IL-5, eotaxin, and IL-13. Journal of Allergy and Clinical Immunology. vol. 111, no. 5, 2003 pp.1049-1061.
- Gamaro, GD., Suyenaga, E., Borsoi, M., Lermen, J., Pereira, P., Ardenghi, P. Effect of rosmarinic and caffeic acids on inflammatory and nociception process in rats. International Scholarly Research Notices. vol.2011. Article ID 451682, 6 pages.
- Ghoshal, A., Das, S., Ghosh, S., Mishra, MK., Sharma, V., Koli, P., Sen, E., Basu, A. Proinflammatory mediators released by activated microglia induces neuronal death in Japanese encephalitis. Glia. vol. 55, no. 5, 2007 pp.483-496.

Giaccone, G. Clinical perspectives on platinum resistance. Drugs. vol. 59, no. 4, 2000 pp. 9-17.

- Halliwell, BARRY., Hoult, JR., Blake, DR. Oxidants, inflammation, and anti-inflammatory drugs. The FASEB journal, vol. 2, no. 13, 1988 pp.2867-2873.
- Harborne, JB., Willam, CA. Advances in flavonoid research since 1992. Phytochemistry. vol. 55, 2000 pp. 481–504.
- Holzmannova, V. Rosmarinic acid and its biological activity. Chemicke listy. vol. 90, no.8, 1996 pp. 486-496.
- Huang, SS., Zheng, RL. Rosmarinic acid inhibits angiogenesis and its mechanism of action in vitro. Cancer Lett. vol. 239, no. 2, 2006 pp. 271-280.
- Ichinose, T., Furuyama, A., Sagai, M. Biological effects of diesel exhaust particles (DEP). II. Acute toxicity of DEP introduced into lung by intratracheal instillation. Toxicology. vol. 99, no. 3, 1995 pp. 153-167.
- Kang, MA., Yun, SY., Won, J. Rosmarinic acid inhibits Ca2+-dependent pathways of T-cell antigen receptormediated signaling by inhibiting the PLC-γ1 and Itk activity. Blood. vol. 101, no.9, 2003 pp 3534-3542.
- Kennedy, DO., Wightman, EL. Herbal extracts and phytochemicals: plant secondary metabolites and the enhancement of human brain function. Advances in Nutrition: An International Review Journal, vol. 2, no.1, 2011 pp. 32-50.
- Kintzios, S., Makri, O., Panagiotopoulos, E., Scapeti, M. In vitro rosmarinic acid accumulation in sweet basil (*Ocimum basilicum* L.). Biotechnol Lett, vol. 25, no.5, 2003 pp. 405-408.
- Kolson, DL., Lavi, E., González-Scarano, F. The effects of human immunodeficiency virus in the central nervous system. Adv. Virus Res. vol. 50, 1998 pp.1-47.
- Kosar, M., Goger, F., Can Başer, KH. In vitro antioxidant properties and phenolic composition of Salvia virgata Jacq. from Turkey. J Agric Food Chem, vol.56, no.7, 2008 pp. 2369-2374.
- Krol, W., Scheller, S., Czuba, Z., Matsuno, T., Zydowicz, G., Shani, J., Mos, M. Inhibition of neutrophils' chemiluminescence by ethanol extract of propolis (EEP) and its phenolic components. Journal of ethnopharmacology. vol.55, no.1, 1996 pp. 19-25.
- Kumagai, Y., Arimoto, T., Shinyashiki, M., Shimojo, N., Nakai, Y., Yoshikawa, T., Sagai, M. Generation of reactive oxygen species during interaction of diesel exhaust particle components with NADPH-cytochrome P450 reductase and involvement of the bioactivation in the DNA damage. *Free Radic. Biol. Med.* vol. 22, no. 3, 1997 pp. 479-487.
- Lamaison, JL., Petitjean-Freytet, C., Carnat, A. Rosmarinic acid, total hydroxycinnamic derivatives and antioxidant activity of Apiaceae, Borraginaceae and

Lamiceae medicinals. In *Annales pharmaceutiques francaises.* vol. 48, no. 2, 1989 pp. 103-108.

- Laufer, S., Greim, C., Bertsche, T. An in-vitro screening assay for the detection of inhibitors of proinflammatory cytokine synthesis: a useful tool for the development of new antiarthritic and disease modifying drugs. Osteoarthritis and cartilage. vol. 10, no.12, 2002 pp. 961-967.
- Lee, J., Jung, E., Kim, Y., Lee, J., Park, J., Hong, S., Hyun, CG., Park, D., Kim, YS. Rosmarinic acid as a downstream inhibitor of IKK- β in TNF- α -induced upregulation of CCL11 and CCR3. Br J Pharmacol. vol. 148, no.3, 2006 pp. 366-375.
- Lee, MY., Lee, NH., Jung, D., Lee, JA., Seo, CS., Lee, H., Kim, JH., Shin, HK. Protective effects of allantoin against ovalbumin (OVA)-induced lung inflammation in a murine model of asthma. Int Immunopharmacol. vol. 10, no. 4, 2010 pp. 474-480.
- Leyck, S., Etschenberg, E., Hadding, U., Winkelmann, J. A new model of acute inflammation: Cobra venom factor induced paw oedema. Inflammation Research. vol.13, no.5, 1983 pp. 437-438.
- Lim, HB., Ichinose, T., Miyabara, Y., Takano, H., Kumagai, Y., Shimojyo, N., Devalia, JL., & Sagai, M. Involvement of superoxide and nitric oxide on airway inflammation and hyperresponsiveness induced by diesel exhaust particles in mice. *Free Radic. Biol* .*Med.* vol. 25, no. 6, 1998 pp. 635-644.
- Mahesh, B., Sathish, S. Antimicrobial Activity of some important medicinal plant against plant and Human Pathogens. World J AgriSci. vol. 4, no.(supp I), 2008 pp.839-843.
- McKay, DL., Blumberg, JB. A review of the bioactivity and potential health benefits of peppermint tea (*Mentha piperita* L.). Phytother Res, vol. 20, no.8, 2006 pp. 619-633.
- Meehye, K. The effect of Prunella on anti-inflammatory activity in RAW264. 7 mouse macrophage cells. Food and Nutrition Sciences 2012. vol.3, 2012.
- Miller, RP., Tadagavadi, RK., Ramesh, G., Reeves, WB. Mechanisms of cisplatin nephrotoxicity. Toxins. vol. 2, no. 11, 2010 pp. 2490-2518.
- Mirzoeva, OK., Calder, PC. The effect of propolis and its components on eicosanoid production during the inflammatory response. Prostaglandins, Leukotrienes and Essential Fatty Acids. vol. 55, no.6, 1996 pp. 441-449.
- Nogueira-Machado, JA., Volpe, CMDO., Veloso, CA., Chaves, MM. HMGB1, TLR and RAGE: a functional tripod that leads to diabetic inflammation. *Expert Opin Ther Targets.* vol. 15, no. 8, 2011 pp. 1023-1035.

Okada, H., Murakami, S. Cytokine expression in periodontal health and disease. Critical Reviews in Oral Biology & Medicine, vol. 9, no.3, 1998 pp.248-266.

- Ola, SS., Catia, G., Marzia, I., Francesco, VF., Afolabi, AA., Nadia, M. HPLC/DAD/MS characterisation and analysis of flavonoids and cynnamoil derivatives in four Nigerian green-leafy vegetables. Food Chem. vol. 115, no. 4, 2009 pp. 1568-1574.
- Osakabe, N., Yasuda, A., Natsume, M., & Yoshikawa, T. Rosmarinic acid inhibits epidermal inflammatory responses: anticarcinogenic effect of *Perilla frutescens* extract in the murine two-stage skin model. Carcinogenesis, vol. 25, no.4, 2004 pp.549-557.
- Osakabe, N., Yasuda, A., Natsume, M., Sanbongi, C., Kato, Y., Osawa, T., Yoshikawa, T. Rosmarinic acid, a major polyphenolic component of *Perilla frutescens*, reduces lipopolysaccharide (LPS)-induced liver injury in D-galactosamine (D-GalN)-sensitized mice. Free Radic Biol Med, vol.33, no.6, 2002 pp. 798-806
- Parnham, MJ., Kesselring, K., Rosmarinic acid. Drugs of the Future, vol. 10, no. 9, 1985 pp. 756–757.
- Patel, R., Attur, MG., Dave, M., Abramson, SB., Amin, AR. Regulation of cytosolic COX-2 and prostaglandin E2 production by nitric oxide in activated murine macrophages. The Journal of Immunology. vol. 162, no. 7, 1999 pp. 4191-4197.
- Peake, PW., Pussell, BA., Martyn, P., Timmermans, V., Charlesworth, JA. The inhibitory effect of rosmarinic acid on complement involves the C5 convertase. International journal of immunopharmacology. vol. 13, no.7, 1991 pp. 853-857.
- Petersen, M. Coleus spp.: in vitro culture and the production of forskolin and rosmarinic acid. Medicinal and Aromatic Plants VI. Springer Berlin Heidelberg, 1994. pp.69-92.
- Petersen, M., Simmonds, MSJ. Rosmarinic acid. Phytochemistry. vol. 62, no.2, 2003 pp. 121- 125.
- Pilotto, A., Sancarlo, D., Addante, F., Scarcelli, C., Franceschi, M. Non-steroidal anti-inflammatory drug use in the elderly. Surgical oncolog. vol. 19, no.3, 2010 pp. 167-172.
- Ramalho, LNZ., Pasta, AAC., Terra, VA., Augusto, MJ., Sanches, SC., Souza-Neto, FP., Cecchini, R., Gulin, F., Ramalho, FS. Rosmarinic acid attenuates hepatic ischemia and reperfusion injury in rats. Food Chem Toxicol. vol. 74, 2014 pp. 270-278.
- Rampart, M., Beetens, JR., Bult, H., Herman, AG., Parnham, MJ., Winkelmann, J. Complementdependent stimulation of prostacyclin biosynthesis: inhibition by rosmarinic acid. Biochemical pharmacology. vol.35, no. 8, 1986 pp. 1397-1400.
- Ravi, V., Parida, S., Desai, A., Chandramuki, A., Gourie-Devi, M., Grau, GE. Correlation of tumor necrosis fac-

tor levels in the serum and cerebrospinal fluid with clinical outcome in Japanese encephalitis patients. J. Med. Virol. vol. 51, no.2, 1997 pp.132-136.

- Rocha, J., Eduardo-Figueira, M., Barateiro, A., Fernandes, A., Brites, D., Bronze, R., Duarte, C.M., Serra, A.T., Pinto, R., Freitas, M. Fernandes, E., Silvia-Lima, B., Mota-Filipe, H., Sepodes, B. Anti-inflammatory Effect of Rosmarinic Acid and an Extract of *Rosmarinus officinalis* in Rat Models of Local and Systemic Inflammation. Basic Clin Pharmacol Toxicol. vol.116, no.5, 2015 pp. 398-413.
- Sagai, M., Furuyama, A., Ichinose, T. Biological effects of diesel exhaust particles (DEP). III. Pathogenesis of asthma like symptoms in mice. Free Radical Biology and Medicine. vol. 21, no. 2, 1996 pp. 199-209.
- Sagai, M., Saito, H., Ichinose, T., Kodama, M., Mori, Y. Biological effects of diesel exhaust particles. I. In vitro production of superoxide and in vivo toxicity in mouse. *Free Radic. Biol. Med.* vol. 14, no. 1, 1993 pp.37-47.
- Sahu, A., Rawal, N., Pangburn, MK. Inhibition of complement by covalent attachment of rosmarinic acid to activated C3b. Biochem Pharmacol. vol. 57, no.12, 1999 pp.1439-1446.
- Salvi, S., Blomberg, A., Rudell, B., Kelly, F., Sandstrom, T., Holgate, S. T., Frew, A. Acute inflammatory responses in the airways and peripheral blood after short-term exposure to diesel exhaust in healthy human volunteers. *Am. J. Respir. Crit. CareMed.* vol. 159, no.(3), 1999 pp. 702-709.
- Sanbongi, C., Takano, H., Osakabe, N., Sasa, N., Natsume, M., Yanagisawa, R., Inoue, K.I., Kato, Y., Osawa, T. Yoshikawa, T. Rosmarinic acid inhibits lung injury induced by diesel exhaust particles. Free Radical Biology and Medicine. vol.34, no.8, 2003 pp. 1060-1069.
- Singh, A., Kulshreshtha, R., Mathur, A. Secretion of the chemokine interleukin-8 during Japanese encephalitis virus infection. J. Med. Microbiol. vol. 49, no. 7, 2000 pp.607-612.
- Sobota, R., Szwed, M., Kasza, A., Bugno, M., Kordula, T. Parthenolide inhibits activation of signal transducers and activators of transcription (STATs) induced by cytokines of the IL-6 family. Biochemical and biophysical research communications. vol. 267, no. 1, 2000 pp. 329-333.
- Sotnikova, R., Okruhlicova, L., Vlkovicova, J., Navarova, J., Gajdacova, B., Pivackova, L., Fialova, S., Krenek, P. Rosmarinic acid administration attenuates diabetesinduced vascular dysfunction of the rat aorta. J Pharm Pharmacol. vol.65, no.5 2013 pp. 713-23.
- Swarup, V., Ghosh, J., Ghosh, S., Saxena, A., Basu, A. Antiviral and anti-inflammatory effects of rosmarinic acid in an experimental murine model of Japanese

encephalitis. *Antimicrob. Agents Chemother*. vol. 51, no.9, 2007 pp. 3367-3370.

- Takano, H., Osakabe, N., Sanbongi, C., Yanagisawa, R., Inoue, K. I., Yasuda, A., Natsume, M., Baba, S., Ichiishi, E.I., Yoshikawa, T. Extract of Perilla frutescens enriched for rosmarinic acid, a polyphenolic phytochemical, inhibits seasonal allergic rhinoconjunctivitis in humans. Exp Biol Med. vol. 229, no. 3, 2004 pp. 247-254.
- Takano, H., Yanagisawa, R., Ichinose, T., Sadakane, K., Inoue, K. I., Yoshida, S. I., Takeda, K., Yoshino, S., Yoshikawa, T., Morita, M. Lung expression of cytochrome P450 1A1 as a possible biomarker of exposure to diesel exhaust particles. *Arch.Toxicol.* vol. 76, no. 3, 2002 pp.146-151.
- Tomlinson, A., Appleton, I., Moore, A. R., Gilroy, D. W., Willis, D., Mitchell, J. A., Willoughby, D. A. Cyclooxygenase and nitric oxide synthase isoforms in rat carrageenin-induced pleurisy. British journal of pharmacology, vol. 113, no.3, 1994 pp. 693-698.
- Upadhyay, S., Dixit, M. Role of polyphenols and other phytochemicals on molecular signaling. Oxidative medicine and cellular longevity. vol. 2015, 2015 Article ID 504253, 15 pages
- Zdarilova, A., Svobodova, A., Simanek, V., Ulrichova, J. Prunella vulgaris extract and rosmarinic acid suppress lipopolysaccharide-induced alteration in human gingival fibroblasts. Toxicology in vitro, vol. 23, no. 3, 2009 pp.386-392.