REVIEW ARTICLE



INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACEUTICAL SCIENCES

Published by JK Welfare & Pharmascope Foundation

Journal Home Page: https://ijrps.com

Applications of Microparticles: A Review

Adiba Khan^{*}, Mitali Bodhankar, Nikita Pal

Department of Pharmaceutics, Gurunanak College of Pharmacy, Nagpur, Maharashtra, India

Article History:	ABSTRACT
Received on: 31 Jan 2023 Revised on: 27 Feb 2023 Accepted on: 01 Mar 2023 <i>Keywords:</i>	The small size and effective carrier capacity of microspheres make them a crucial component of novel drug delivery systems. With a particle size range of 1-1000 μ m, microspheres are distinctively free-flowing powders made of synthetic polymers or proteins. The range of microsphere production methods provides several chances to regulate drug delivery processes and improve the
Microspheres,	therapeutic potency of certain medications. There are numerous techniques
Target Site,	for delivering therapeutic medicines to target areas over time. One such tech-
Controlled Release,	nique is to employ microspheres, also known as microparticles, as medication
Novel Drug Delivery	carriers. When adjusted and kept at the correct concentrations at the location of interest, it provides a dependable method of delivering medications selec- tively to target areas. In addition to their capabilities for prolonged release, microspheres have received a lot of attention.

^{*}Corresponding Author

Name: Adiba Khan Phone: 9527410706 Email: adiba8423@gmail.com

ISSN: 0975-7538

DOI: <u>https://doi.org/10.26452/ijrps.v14i1.4256</u>

Production and Hosted by

IJRPS | https://ijrps.com

© 2023 | All rights reserved.

INTRODUCTION

Microspheres characteristically are free flowing powders which consist of natural/synthetic polymers and diameters having in the range of 1–1000 μ m. Microspheres are the microparticulate carrier system which is used nowadays as novel drug delivery approach. The therapeutic substance is delivered to the target location by the microspheres at a pace and concentration that allows for maximum therapeutic efficacy while minimizing adverse effects. It does, however, have several shortcomings. Microspheres denature within a few weeks, making them relatively unstable and not easy to formulate [1].

Multiparticulate carriers are used for site specific drug delivery. Solid microspheres nowadays used for various applications [2]. Many newer drugs face problems such as low solubility, high potency with poor stability; the drug delivery of these drugs impacts efficacy and potential for commercialization and nature of drug. As a result, there are a number of drug delivery techniques that deliver the therapeutic agent in the appropriate quantity and at the right time to the right area in the body, maximizing efficacy and minimizing adverse effects while also increasing compliance.

The formulation of microspheres has been studied using a variety of materials, both biodegradable and non-biodegradable. These materials include polymers with natural and synthetic origins as well as naturally occurring compounds that have been altered. Table 1 classifies and provides a list of some of the polymers utilized in the creation of microspheres.

Multiparticulate Carriers in Various Drug Delivery Applications

Microspheres in Vaccine Delivery

Chronic autoimmune inflammation of the pancreatic cells producing insulin results in type 1 diabetes, a condition of glucose imbalance [3, 4]. The final result is a deficiency in insulin-producing cells,

Syntheti	Natural Materials			
Non-biodegradable	Biodegradable	Proteins	Carbohydrates	Chemically mod- ified carbohy- drates
Poly(methyl methacrylate)	Polyphosphazenes	Albumins	Starch	Poly(acryl)dextran
Acrolein	Polyalkyl cyanoacry- lates	Gelatin	Agarose	Poly(acryl)starch
Glycidyl methacrylate Epoxy polymers	Polyanhydrides	Collagen	Carrageenan Chitosan	

Table 1: Classification of Polymers

defined as the number of cells below a threshold that is crucially needed to maintain physiological glucoregulation. Insulitis and peri-insulitis, two growing infections surrounding and inside the pancreatic cells, first render the insulin-producing cells insensitive to glucose and incapable of releasing the proper amount of insulin, mostly as a result of the activities of cytokines including interferon, tumour necrosis factor, and interleukin [3, 5, 6].

Many type 1 diabetics still exhibit signs of a residual β -cell mass that functionally reacts to glucose and generates insulin for a short period of time after their condition is clinically established [3, 7].

Patients who still had β -cell masses actually had good glycemic control and a better prognosis for diabetes consequences including retinopathy and nephropathy. These findings have compelled researchers to look at possible medications that may be utilized to prevent the loss of any residual betacell mass at clinical diagnosis, particularly by preventing active autoimmunity. Autoimmunity might be controlled initially with the use of pharmacological systemic immunosuppressive drugs, but once they were stopped, the condition returned, requiring long-term administration of systemic drugs with their accompanying adverse effects [3, 8, 9].

While clinical withdrawal of hyperglycemia has recently been accomplished with the infusion of anti-CD3 antibodies, problems remain surrounding the mechanism of action of momentary immunodepletion and its associated cytokine-related consequences [10, 11].

It is widely known that the antigen-presenting cells that infiltrate pancreatic islets in response to as-yetunidentified microenvironmental alterations make up the majority of the initial wave of infiltrating immune cells during the immunopathogenesis of type 1 diabetes [12].

This issue results in the chronic process of migrating antigen-presenting cells, specifically dendritic cells, being formed from apoptotic and/or necrotic cells, although not being fully resolved mechanistically and chronologically. An endogenous "maturation programme" that accumulates in pancreatic lymph nodes and activates T cells, including autoreactive T cells that are unique to β -cells, occurs in migrating dendritic cells [13–15].

Additionally, immunomodulatory "suppressive" cellular networks can be activated and maintained by dendritic cells. Exogenous infusion of functionally immature dendritic cells has been shown to increase allograft survival and avoid autoimmune illness and its recurrence, according to several studies [16]. Syngeneic recipients received dendritic cells obtained from NOD mice that had low expression of CD40, CD80, and CD86 (produced by ex vivo treatment with antisense oligonucleotides targeted to the 5' end of each main transcript). It has been transfected, and it greatly delays and prevents the beginning of disease [17].

Many studies have demonstrated that multiparticulate carriers may direct dendritic cells to the administration site and that post-phagocytic contents can influence the functional expression of dendritic cells [18].

Biodegradable poly(lactic-co-glycolic acid) (PLGA) microspheres have been shown by Yoshida and Babensee [3, 18] to either promote dendritic cell senescence by activating the co-stimulatory molecules CD40, CD80, and CD86. These experiments employed a nucleic acid delivery mechanism that dendritic cells could phagocytize without up-regulating these co-stimulatory molecules [3]. The PROMAXX microsphere delivery method was therefore modified to include antisense oligonucleotides direct against CD40, CD80, and CD86 [3]. More prominently, as compared to the known immunomodulating features of PLGA-based formulations when delivered in vivo, this method is neutral with regard to the maturation status of

dendritic cells [3].

When designing microsphere chemistry for immunosuppressive targets that use dendritic cells as mediators, one crucial criterion is the neutrality of dendritic cell development. This paper describes a PROMAXX microsphere-based vaccination where it has been demonstrated that antisense oligonucleotides can inhibit diabetic dendritic cells and stop or even revert the onset of autoimmune diabetes [16].

Microspheres in Ocular Delivery Systems

The fast elimination of traditional liquid eye drops from eyes remains one of the key issues with eye drop administration. A high tear drainage rate is caused by a variety of causes, including fast tear turnover and associated pre-corneal loss, production of tear outflow by irritant formulations, and delivery of relatively large eye drops. The drug half-lives in the cornea of various pharmacological formulations are between one and three minutes. Because of this, very little of the dosage, only around 1–3% of the total dose, may actually pass through the cornea and reach the intraocular tissues [19, 20]. A considerable proportion of the medicine is instead expelled in the nose and intestines due to inadequate productive absorption. Particularly the nose and the intestines are particularly effective bodily organs for absorption. This may cause a significant amount of systemic absorption, which may result in undesirable side effects and medication toxicity [20]. These issues have been recognized for some time, but relatively little has been done by pharmaceutical companies to address them, and there aren't many other options for delivering ophthalmic drugs on the market [3].

The utilisation of microparticles is one option for such systems. The benefit of these colloidal particles is that they may be used as eye drops or other liquid applications. So, the discomfort related to applying viscous or sticky substances, such as ointments, may be avoided. If the prior preparations are used correctly, they cause complete visual blur. Even when manufactured as non-dissolving inserts, it might be challenging for older patients to remove them after administration of a big insert.

Liposomes are yet another option for micro particle systems. Liposomes, on the other hand, may be less stable. Microspheres and microcapsules, as well as nanoparticles and nanocapsules, are examples of suitable micro particle systems.

The particle sizes of microparticles and microcapsules are higher than 1 μ m. Microcapsules are made up of a polymer membrane enclosing a solid or liquid drug reservoir, while microspheres are monolithic particles with a porous or solid polymer matrix. The medication is either suspended in the form of a solid dispersion or is dissolved in the polymer matrix as a solid solution in monolithic microparticles. The ocular bioavailability of these particles is significantly higher than that of conventional aqueous ophthalmic solutions after appropriate drug conjugation to them [20].

Choroidal neovascularization is the disease occurring in developing countries. Various anti-VEGF medicines have received approval from the US FDA for the treatment of this disease. The current norm for giving anti-VEGF treatments is monthly (or biweekly) intravitreal bolus injections. Serious side effects such endophthalmitis, retinal detachment, intravitreal haemorrhage, and cataracts might result after injectable administration. The use of controlled release microspheres lowers the likelihood of problems. The delivery composition consists of microcapsules suspended in a thermoresponsive hydrogel, such as nano- or microspheres or other similar microencapsulated structures. The medicinal substances are released in a regulated manner by the microencapsulated particles [21].

Microspheres as Nasal Drug Delivery Systems

Protein and peptide medication delivery is accomplished via the intranasal route. The nasal mucosa quickly clears the traditional dosing formulations. It has been suggested that using bioadhesive gels can improve insulin and calcitonin retention. The gel dose formulations are replaced with the bioadhesive microspheres. The surface characteristics and release pattern of the bioadhesive microspheres can be more precisely controlled than they can with the gel dosage form.

The lipophilic nature of intranasal drug delivery microspheres allows for some water absorption, which causes the spheres to expand and create a gel. Starch, albumin, hyaluronic acid, dextran, and bioavailability of proteins and peptides were increased in several animal models in the composition of microspheres used for nasal medication administration. The development of microspheres also uses medications with low molecular weight. By physically interacting with the nasal mucosa, these microspheres cause epithelial cell tight junctions to open. Dextran and starch microspheres were employed as an efficient dose for nasal medication administration [22].

Magnetic Microspheres

A novel method for achieving site-specific medication delivery is to target drugs under controlled,

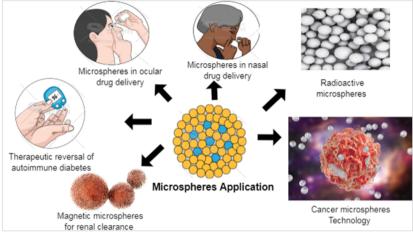


Figure 1: Application of Microspheres

-			
Inventor Name and Year	Patent Title	Patent No.	Ref.
Pankaj Modi et al. (1996)	Vaccine delivery system for immunization, using biodegrad- able polymer microspheres	US5569468A	[23]
Hyeon-Kook Lee et al. (1998)	Single-Shot vaccine Formulation	US5753234A	[24]
Hamilton et al (1998)	Drugs, vaccines and hormones in polylactide coated microspheres	EP0744940B1	[25]
P. Andrew Lennard et al. (2014)	Microspheres for the treatment of brain tumors	US008691791B2	[26]
J. Kang-Mieler et al (2017)	Biodegradable microsphere- hydrogel ocular drug delivery system	US20170087248A1	[21]

Table 2: Patents on Microparticles

burst, or modulated release. It is useful to look at how the body places its own biopharmaceuticals in the targeted tissues when creating new methods of targeting. The benefit of magnetic monitoring is that it effectively enables high local concentrations of medicinal drugs. This is the exact usage of magnetic medication delivery with particle carriers. When therapeutic drugs are injected into a patient's circulation and halted at the target spot by a strong magnetic field, radioisotopes are employed for magnetic monitoring [27].

Medication targeting refers to the exclusive delivery of a drug to receptors, organs, or other targeted areas of the body. Although magnetic microspheres have been used successfully to target drugs, they are not very site-specific and are often quickly eliminated by the reticuloendothelial system. Magnetic microspheres are made to promote targeting of active drugs while reducing clearance. They can be utilised to catch different medicines.

Due to its significantly higher phagocytic activity than normal cells in the event of solid tumour targeting, magnetic microsphere can infiltrate tumour cells [3].

Cancer Microsphere Technology

How to combat cancer on a molecular level has been learned as a result of the research of microspheres. SIR-Spheres microspheres are radioactive polymeric spheres that generate beta radiation. Large amounts of microspheres are delivered immediately to the cancer cells once it has been injected into the hepatic artery using a catheter. Microspheres made of SIR-Spheres are used to attack liver cancers and shield healthy liver tissue [3].

Normal traditional dose forms for cancer therapy kill normal cells in addition to malignant cells. Microspheres are therefore employed to target tumour tissues while without having an adverse effect on healthy cells.

The main brain tumour known as glioblastoma multiforme, also called grade 4 astrocytoma, accounts for 52% of all primary brain tumours and 20% of intracranial tumours. Palliative care includes surgical procedures, chemotherapy, and radiation therapy. A variety of brain cancers are treated using microspheres as well [28].

Radioactive Microspheres

When the encapsulated diagnostic radioisotopes are substituted with medicinal radioisotopes from the group of α - or β -emitters, radio labelled microspheres are acceptable for therapy. Topical application is frequently used to treat conditions including rheumatoid arthritis, liver tumours, and cystic brain tumours.

However, because of radiochemical instability and inadequate biodistribution of the radiopharmaceutical entity, its utility is limited by unintended toxicity, lower than anticipated target absorption, and poor therapeutic effectiveness. Despite the apparent superiority of many radiation therapies, there remains a common aversion to using radioactive materials [3, 29].

Perfect Count Microspheres

For use in in vitro diagnosis, these microspheres are available. These microspheres are designed to be used with flow cytometry to count cells absolutely in samples of medium, bone marrow, peripheral blood, and leukophoresis.

These microbead-based, single-platform methods can assist identify the cell subpopulations for which absolute counting is desired by combining them with monoclonal antibodies attached to various fluorochromes [Figure 1] [3, 30].

Patents Based on Microparticles

The Patents Based on Microparticles is shown in Table 2.

CONCLUSION

The availability of a variety of biodegradable and bio-inspired polymers has enabled extended drug release, effective immunization, and in the development of bioimplantable, bioreactors, biochips. It is now possible to design microsphere systems for the development of biosensors and tissue replacement. Microspheres will play a key role in innovative drug delivery systems in the future, notably in diagnostics. For kidney stone detection Retrograde Pyelogram used as dye which is injected through catheter and X-rays are taken. Without injecting this dye we can use microspheres coated with this dye taken orally can detect the kidney stone without injecting or anesthetizing. Especially in the fields of diagnosis, organ imaging, genetics, and drug development, the future of microspheres is undoubtedly promising.

Funding Support

The authors declare that they have no funding support for this study.

Conflict of Interest

The authors declare that there is no conflict of interest.

REFERENCES

- [1] M K Das, A B Ahmed, and D Saha. Microsphere a drug delivery system: A review. *Int J Curr Pharm Res*, 11(4):34–41, 2019.
- [2] P Parida, S C Mishra, S Sahoo, A Behera, and B P Nayak. Development and characterization of ethylcellulose based microsphere for sustained release of nifedipine. *Journal of pharmaceutical analysis*, 6(5):341–344, 2016.
- [3] K J Beyatricks. Recent trends in microsphere drug delivery system and its therapeutic applications A Review. *Crit. Rev. Pharm. Sci*, 2(1):1–14, 2013.
- [4] M A Atkinson and G S Eisenbarth. Type 1 diabetes: new perspectives on disease pathogenesis and treatment. *The Lancet*, 358(9277):221–229, 2001.
- [5] H E Thomas, R Darwiche, J A Corbett, and T W Kay. Interleukin-1 plus γ -interferoninduced pancreatic β -cell dysfunction is mediated by β -cell nitric oxide production. *Diabetes*, 51(2):311–316, 2002.
- [6] M Arnush, M R Heitmeier, A L Scarim, M H Marino, P T Manning, and J A Corbett. IL-1 produced and released endogenously within human islets inhibits beta cell function. *The Journal of clinical investigation*, 102(3):516– 526, 1998.
- [7] M Abdul-Rasoul, H Habib, and M Al-Khouly.
 'The honeymoon phase' in children with type 1 diabetes mellitus: frequency, duration, and influential factors. *Pediatric diabetes*, 7(2):101–107, 2006.
- [8] P F Bougnères, P Landais, C Boisson, J C Carel, N Frament, C Boitard, and J F Bach. Limited duration of remission of insulin dependency in children with recent overt type I diabetes treated with low-dose cyclosporin. *Diabetes*, 39(10):1264–1272, 1990.
- [9] R Lipton, R E Laporte, D J Becker, J S Dorman, T J Orchard, J Atchison, and A L Drash. Cyclosporin therapy for prevention and cure of IDDM: epidemiological perspective of benefits and risks. *Diabetes care*, 13(7):776–784, 1990.

- [10] L Chatenoud. CD3-specific antibodies restore self-tolerance: mechanisms and clinical applications. *Current opinion in immunology*, 17(6):632–637, 2005.
- [11] K C Herold, S E Gitelman, U Masharani, W Hagopian, B Bisikirska, D Donaldson, and J A Bluestone. A single course of anti-CD3 monoclonal antibody hOKT3 γ 1 (Ala-Ala) results in improvement in C-peptide responses and clinical parameters for at least 2 years after onset of type 1 diabetes. *Diabetes*, 54(6):1763–1769, 2005.
- [12] N Babaya, M Nakayama, and G S Eisenbarth. The stages of type 1A diabetes. *Annals of the New York Academy of Sciences*, 1051(1):194– 204, 2005.
- [13] R S Allan, J Waithman, S Bedoui, C M Jones, J A Villadangos, Y Zhan, and F R Carbone. Migratory dendritic cells transfer antigen to a lymph node-resident dendritic cell population for efficient CTL priming. *Immunity*, 25(1):153–162, 2006.
- [14] F R Carbone, G T Belz, and W R Heath. Transfer of antigen between migrating and lymph noderesident DCs in peripheral T-cell tolerance and immunity. *Trends in immunology*, 25(12):655– 658, 2004.
- [15] C Scheinecker, R Mchugh, E M Shevach, and R N Germain. Constitutive presentation of a natural tissue autoantigen exclusively by dendritic cells in the draining lymph node. *The Journal of experimental medicine*, 196(8):1079–1090, 2002.
- [16] K R Mccurry, B L Colvin, A F Zahorchak, and A W Thomson. Regulatory dendritic cell therapy in organ transplantation. *Transplant international*, 19(7):525–538, 2006.
- [17] J Machen, J O Harnaha, R Lakomy, A Styche, M Trucco, and N Giannoukakis. Antisense oligonucleotides down-regulating costimulation confer diabetes-preventive properties to nonobese diabetic mouse dendritic cells. *The Journal of Immunology*, 173(7):4331–4341, 2004.
- [18] M Yoshida and J E Babensee. Molecular aspects of microparticle phagocytosis by dendritic cells. *Journal of Biomaterials Science*, 17(8):893–907, 2006.
- [19] V H L Lee and J R Robinson. Mechanistic and quantitative evaluation of precorneal pilocarpine disposition in albino rabbits. *Journal of pharmaceutical sciences*, 68(6):673–684, 1979.

- [20] A Zimmer and J Kreuter. Microspheres and nanoparticles used in ocular delivery systems. *Advanced drug delivery reviews*, 16:61– 73, 1995.
- [21] J Kang-Mieler and E Brey. Biodegradable microsphere hydrogel ocular drug delivery system, 2017. United States Patent No. US 2017/0087248 A1.
- [22] L Pereswetoff-Morath. Microspheres as nasal drug delivery systems. *Advanced Drug Delivery Reviews*, 29(1-2):185–194, 1998.
- [23] P Modi. Vaccine Delivery System For Immunization, Using Biodegradable Polymer Microspheres 76, 1996. USO05569468A.
- [24] H K Lee, J H Park, N S Choi, M J Kim, and S H Kim. U.S. Patent No. 5,753,234. Washington, DC: U.S. Patent and Trademark Office, 1998.
- [25] Hamilton. Drugs, vaccines and hormones in polylactide coated microspheres, 2019. EP0744940B1.
- [26] P W S Andrew Lennard Lewis and Yiqing Tang. Microspheres for treatment of brain tumors. United States Patent No. US 8,691,791 B2.
- [27] U O Häfeli. Magnetically modulated therapeutic systems. *International journal of pharmaceutics*, 277(1-2):19–24, 2004.
- [28] M S Rajput and P Agrawal. Microspheres in cancer therapy. *Indian Journal of Cancer*, 47(4):458–468, 2010.
- [29] P G Rose, B N Bundy, E B Watkins, J T Thigpen, G Deppe, M A Maiman, and S Insalaco. Concurrent cisplatin-based radiotherapy and chemotherapy for locally advanced cervical cancer. *New England Journal of Medicine*, 340(15):1144–1153, 1999.
- [30] B Brando, D Barnett, G Janossy, F Mandy, B Autran, and G Rothe. Cytofluorometric methods for assessing absolute numbers of cell subsets in blood. *Cytometry*, 42(6):327–346, 2000.