ORIGINAL ARTICLE



INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACEUTICAL SCIENCES

Published by JK Welfare & Pharmascope Foundation

Journal Home Page: <u>https://ijrps.com</u>

Identify the Interleukin-6 gene -597 A/G Polymorphism in Type 2 diabetic patients

Haider Abd Jabbar Alammar*1, Sahar Dakhil Yonis2, Zainab A Razak Al-Sharifi3

¹Department of Biochemistry, College of Medicine, University of Al-Qadisiyah, Iraq ²Department of Anatomy, College of Medicine, University of Al-Qadisiyah, Iraq ³Department of Biochemistry, College of Medicine, University of Baghdad, Iraq

Article History:	ABSTRACT
Received on: 04.01.2018 Revised on: 23.04.2018 Accepted on: 24.04.2018	Tumor necrosis factor-alpha (TNFa), and interleukin-6 (IL-6) are a cytokine which has a role in a metabolic progression which might facilitate diabetes progression. The current study was aimed to investigate the presence of Interleukin-6 gene -597 A/G Polymorphism in blood samples of 64 diabetic and
Keywords:	30 blood samples healthy control in Al-Diwaniyah City, Iraq. Using PCR RFLP, the results have shown that IL-6 polymorphism was pronounced in the pa-
Arthroscopy, Long-term results, Partial meniscectomy	tients with type 2 diabetes, particularly the genotype AA, GG, and AG and it showed s significant association with sex and age parameter. Also, this study found that the prevalence (Carriage rates) of allele A was is much lower than that in (Carriage rates) allele G in T2DM population compared to healthy pop- ulation, which is confirmed the previous results. The current data will pro- vide a sight on the role of heterogeneity of cytokine gene in the development of T2DM. Furthermore, these cytokine genes in addition of its variants might be a potential indicator for diseases susceptibility in south Iraq.

* Corresponding Author

Name: Haider Abd Jabbar Alammar Phone: +9647818281147 Email: habd959@gmail.com

ISSN: 0975-7538

DOI: https://doi.org/10.26452/ijrps.v9i2.1482

Production and Hosted by

IJRPS | <u>https://ijrps.com</u> © 2018 | All rights reserved.

INTRODUCTION

Diabetes is one of the chronic metabolic disease, which might lead to a high morbidity and mortality in adults worldwide. In 2013 only, nearly 382.000.000 people were diagnosed with diabetes mellitus (T2DM),and this number might arise to 592.000.000 by 2035 (Guariguata *et al.*, 2014). Type 2 diabetes mellitus is the most form of diabetes with around 90 % of all diagnostic cases. Over earlier decades, diabetes has transformed from a mild disease to one of the major diseases that affecting both the young and adults. In addition, the

Type 2 diabetes mellitus (T2DM) was increased from 2.39% in 2000 to 5.32% in 2013 (Sharma et al., 2016, Guariguata et al., 2014, Saxena et al., 2013). White blood cells are highly expressed Interleukins, mainly leukocytes which facilitate immune reactions and play a major role in the pathogenesis of T2DM (Zhuanping et al., 2016). It has been shown that cytokines such as tumor necrosis factor-alpha and interleukin-6 have a role in the metabolic process which might mediate diabetes progression, whereas interleukin-6 (IL-6) and IL-10 impart protection. Polymorphisms in the codon region of these cytokine genes might affect on their structure as well as expression (Saxena et al., 2012, Erbilgin et al., 2013). Studies suggested that T2DM is correlated with a systemic inflammation as results of increase blood level of immune response of both markers and mediators. This due to the association of increase plasma IL-6 levels with patients has T2DM and also with impaired glucose tolerance, which suggests its potential role in the type 2 diabetes progression (Pradhan et al., 2001, MŘller et al., 2002). The high level of IL-6 in plasma is regulated by increasing of transcription and translation at the molecular level which might be affected

by existence of polymorphisms in its coding region (Pakala et al., 1997). A study indicated that the single nucleotide polymorphisms (SNPs) located in IL-6 promoter might increase the risk factor of T2DM disease (Terry et al., 2000). It has been noticed that increase in the expression of TNF-a in the muscle tissues led to decreases tyrosine kinase signaling, suggesting its potential role in diabetes and insulin resistance (Hotamisligil and Spiegelman, 1994, Tsiotra et al., 2001). It was indicated that 75 % of IL-10 secretion is due to genetic process and its secretion is mainly controlled by the gene transcription in the nucleus. A study suggested that a combination of CA, GG and AA for TNF-a, IL-6, and IL-10 gene polymorphisms might lead to increase the susceptibility and the risk of developing T2DM (Saxena et al., 2013). Therefore, this study examined the association of IL-6 gene promoter polymorphism in the association with T2DM development.

PATIENTS AND METHODS

Samples collections

EDTA tubes were used to collect 64 blood samples taken from patients that were diagnosed with type 2 diabetes and 30 blood samples healthy control individual from the diabetic center in Al-Diwaniya Teaching Hospital. The samples were stored in the refrigerator until use for DNA extraction.

Genomic DNA Extraction

This study used a Genomic DNA Mini Kit, Geneaid. USA to extract Genomic DNA from frozen blood samples. The extraction was achieved by using frozen Blood extraction technique with Proteinase K according to company instructions. Next, the DNA concentration was tested by Nanodrop spectrophotometer, and then the samples placed in -20C until use for conventional PCR.

PCR RFLP

Detection of 597 A/G polymorphism in IL-6 gene was achieved using PCR-RFLP techniques from blood samples of T2DM patients and healthy. This producer was accomplished according (Saxena et al., 2014). The primers were purchased from Bioneer company, Korea) with forwarding sequences (5'-GGAGTCACA CACTCCACCT-3) and reverse sequence (5'-CTTAATAAGGTTTCC AATCAG-3). The master mix was prepared according to (AccuPower® PCR PreMix kit. Bioneer. Korea) provided protocol. The PCR tubes contain a pellet which was consist of (Taq DNA polymerase 1U, Tris-HCl (pH 9.0) 10mM, KCl 30mM, stabilizer, MgCl2 1.5mM, dNTPs 250µM, and tracking dye). The preparation of master mix was achieved according to the protocol of provided kit. This achieved by adding 5µl of DNA and 1.5µl of 10pmole of each forward primer and reverse primer mixed together, and then complete the volume to the 20µl of deionizer PCR water. All the cocktail above was mixed, vortexed then placed in the thermocycler (T100 Thermal cycler Biorad. USA) and following protocol was applied; initial denaturation temperature was 94°C for 5 min; followed by 35 cycles at denaturation 94°C for 30 s, annealing was 57°C for 30 s, and extension was 72°C for 1

Table 1: Clinical and controls characteristics & patients cases		
Clinical characteristics	Control(N-20 case)	Pationts (N= 6

14010 1				
Clinical	l characteristics	Control (N= 30 case)	Patients (N= 64 case)	P value
60V	Male	7(23.33%)	27(42.18%)	0.07
sex	Female	23(71.42%)	37(57.81%)	0.07
Age		32.46±2.31	39.81±1.16	0.007*
	a			

Values of age represent mean ± standard error

* significant differences at level 5%

Table 2: Genotype frequencies

Tuno	Groups	
Туре	Controls number & count (%) (N=30)	Cases number & count (N= 64)
AA	13(43.33)	45(70.31)
GG	6(20)	7(10.93)
AG	11(36.66)	12(18.75)
P value	0.042	

Significant differences at level 5%

Table 3: Allele frequencies

Туре	Groups	
	Controls number & count (%) (N=30)	Cases number & count (N= 64)
A	37(61.66)	102(79.68)
G	23(38.33)	26(20.31)
P value	0.009	
Odds ratio (95% CI)	0.4101(0.2087 to 0.8058)	

Туре	Groups	
	Controls number & count (%) (N=30)	Cases number & count (N= 64)
A(+)	24 (80)	57(89.06)
A(-)	6 (20)	7(10.93)
P value	0.241	
Odds ratio (95% CI)	0.4912(0.1494 to 1.6150)	

Table 4: Carriage rates for allele A

Туре	Groups	
	Controls number & count (%) (N=30)	Cases number & count (N= 64)
G(+)	17(43.33)	19(29.68)
G(-)	13 (52.38)	45(70.31)
P value	0.013	
Odds ratio (95% CI)	3.097(1.2599 to 7.6137)	

min and final extension was 72°C for 5 min. an agarose gel with 2% was used to examine the PCR products using ethidium bromide, and then visualized with UV illumination. RFLP techniques was achieved using (FokI, Biolabs, UK) restriction enzyme to detect IL-6 gene 597 A/G polymorphism; (AA) wild-type homozygote, the product digested by restriction enzyme and the band size was 527bp, the (A/G) heterozygote, the product was digested by restriction enzyme and the bands size were 527bp, 461bp, and 66bp, the (GG) mutant type homozygote, was digested by restriction enzyme and the bands size were 461bp and 66bp.

RESULTS

The current study investigated the effects of sex on the genotypes of the IL-6 gene in the patient compared to the healthy group. The results showed that there were significant effects of the gender parameter (p<0.05) on the IL-6 gene polymorphism. The effects of the age parameter on the IL-6 gene polymorphism were also investigated, and the results indicate a significant (p<0.05) increased of the IL-6 gene polymorphism within the age in the patient group compared to healthy group table1.

The genotype frequencies of the IL-6 gene polymorphism were examined in the patient group compared to healthy control group. For the AA, GG and AG genotype, the results showed a significant (p<0.05) presence of these genotypes in the patient group compared to healthy control group. In addition, this study investigated the allele A and Allele G frequencies in the patient population as well as in the healthy population, and the results showed a significant shifting in the both of alleles compared to healthy group table 3. The Carriage rates for allele A and G were examined in the patient population compared to the healthy population, and no significant changes were noticed in Carriage rates for allele Aalleles. However, a significant change was noticed in Carriage Rates G alleles

in the patient population compared to healthy population table 4,5.

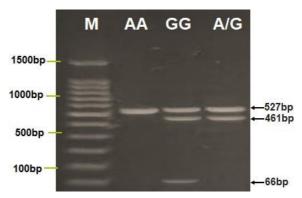


Figure 1: Agarose gel electrophoresis image that shows the RFLP-PCR product analysis of interleukin-6 (*IL-6*)-597 A/G (rs1800797) polymorphism using FokI restriction enzyme. Where M: marker (1500-50bp), The (AA) wildtype homozygote, the product undigested by restriction enzyme 527bp band. The (GG) mutant type homozygote that shows digested by restriction enzyme into 461bp and 66bp band. The (A/G) heterozygote, the product digested by restriction enzyme into 527bp, 461bp and 66bp band.

DISCUSSION

The current study found that Interleukin-6 gene -597 A/G Polymorphism showed significant correlation with T2DM. This study also suggested that SNP -597 which is located in the IL-6 gene promoter region might be a risk factor to develop of T2DM. This might give a suggestion that there is a strong link between T2DM and IL-6 gene polymorphisms which might consider as a prognostic biomarker for progression of type 2 diabetes. The results from this study is reasonable because an early study has demonstrated that in addition to role of IL-6 as immune regulator, it also has a role in glucose homeostasis as well as glucose metabolism inside adipocytes, hepatocytes, skeletal muscle cells, neuroendocrine cells and pancreatic b-cells (Han et al., 2017). In addition, a meta-analysis study has supported our finding which indicated that there is a link between the genetic change of IL-6 gene with increasing risk factor to develop T2DM (Yin et al., 2013). The IL-6 polymorphism was more pronounced in the patients with type 2 diabetes, particularly the genotype AA, GG, and AG and it showed s significant association with sex and age parameter. The females were showed more occurrence of this genotype of the IL-6 gene in their samples with 57.81% percentage. Therefore, IL-6-597 A/G Polymorphism that the frequency of -592 IL-10 -592 C/A polymorphism shown that the frequency A\G alleles was significantly increased in type 2 diabetes patients compared to healthy. This suggesting that this SNP has a crucial role in susceptibility to develop type 2 diabetes, which is agreed with (Chang et al., 2005). Also, this study found that the prevalence (Carriage rates) of allele A (0.4912%) was is much lower than that in (Carriage rates) allele G (3.097%) in T2DM population compared to healthy population, which is confirmed the previous results. This investigational study found that AA genotype increased the risk up to 70.31% in T2DM compared to healthy group 43.33%. While for GG and AG was decreased the risk in the T2DM compared to healthy control. Also, allele A frequency increased to approximately to 79.68% in the T2DM compared to healthy. While, allele G frequency was not given any significant results (Saxena et al., 2013).

CONCLUSION

This finding suggested that that genotypes and alleles are associated with progression of type 2 diabetes, however only in combination. The current data will provide a sight on the role of heterogeneity of cytokine gene in the progression of T2DM. Furthermore, these cytokine genes in addition of its variants might be a potential indicator for diseases susceptibility in south Iraq.

REFERENCES

- Chang, Y.-H., Huang, C.-N., Wu, C.-Y. & Shiau, M.-Y. 2005. Association of Interleukin-10 A-592c and T-819c Polymorphisms with Type 2 Diabetes Mellitus. *Human Immunology*, 66, 1258-1263.
- Erbilgin, A., Civelek, M., Romanoski, C. E., Pan, C., Hagopian, R., Berliner, J. A. & Lusis, A. J. 2013. Identification of Cad Candidate Genes in Gwas Loci and Their Expression in Vascular Cells. *Journal of Lipid Research*, 54, 1894-1905.
- Guariguata, L., Whiting, D. R., Hambleton, I., Beagley, J., Linnenkamp, U. & Shaw, J. E .2014. Global Estimates of Diabetes Prevalence For 2013 And Projections For 2035. *Diabetes Research and Clinical Practice*, 103, 137-149.

- Han, C., Zhang, M., Luo, X., Wang, C., Yin, L., Pang, C., Feng, T., Ren, Y., Wang, B. & Zhang, L. 2017. Secular Trends in The Prevalence of Type 2 Diabetes in Adults in China From 1995 To 2014: A Meta-Analysis. *Journal of Diabetes*, 9, 450-461.
- Hotamisligil, G. S. & Spiegelman, B. M. 1994. Tumor Necrosis Factor A: A Key Component of The Obesity-Diabetes Link. *Diabetes*, 4.1271-1278, 3
- Mřller, S., Martin, S., Koenig, W., Hanifi-Moghaddam, P., Rathmann, W., Haastert, B., Giani, G., Illig, T., Thorand, B. & Kolb, H. 2002. Impaired Glucose Tolerance Is Associated with Increased Serum Concentrations of Interleukin 6 And Co-Regulated Acute-Phase Proteins but Not Tnf-A Or Its Receptors. *Diabetologia*, 45, 805-812.
- Pakala, S. V., Kurrer, M. O. & Katz, J. D. 1997. T Helper 2 (Th2) T Cells Induce Acute Pancreatitis and Diabetes in Immune-Compromised Nonobese Diabetic (Nod) Mice. *Journal of Experimental Medicine*, 186, 299-306.
- Pradhan, A. D., Manson, J. E., Rifai, N., Buring, J. E. & Ridker, P. M. 2001. C-Reactive Protein, Interleukin 6, And Risk of Developing Type 2 Diabetes Mellitus. *Jama*, 286, 327-334.
- Saxena, M., Agrawal, C. G., Bid, H. K. & Banerjee, M. 2012. An Interleukin-10 Gene Promoter Polymorphism (– 592a/C) Associated with Type 2 Diabetes: A North Indian Study. *Biochemical Genetics*, 50, 549-559.
- Saxena, M., Srivastava, N. & Banerjee, M. 2013. Association of Il-6, Tnf-A And Il-10 Gene Polymorphisms with Type 2 Diabetes Mellitus. *Molecular Biology Reports*, 40, 6271-6279.
- Sharma, M., Nazareth, I. & Petersen, I. 2016. Trends in Incidence, Prevalence and Prescribing in Type 2 Diabetes Mellitus Between 2000 And 2013 In Primary Care: A Retrospective Cohort Study. *Bmj Open*, 6, E010210.
- Terry, C. F., Loukaci, V. & Green, F. R. 2000. Cooperative Influence of Genetic Polymorphisms on Interleukin 6 Transcriptional Regulation. *Journal of Biological Chemistry*, 275, 18138-18144.
- Tsiotra, P. C., Tsigos, C. & Raptis, S. A. 2001. Tnfα And Leptin Inhibit Basal and Glucose-Stimulated Insulin Secretion and Gene Transcription in The Hit-T15 Pancreatic Cells. *International Journal of Obesity*, 25, 1018.
- Yin, Y. W., Sun, Q. Q., Zhang, B. B., Hu ,A. M., Liu, H. L., Wang, Q., Zeng, Y. H., Xu, R. J., Zhang, Z. D. & Zhang, Z. G. 2013. Association Between the Interleukin-6 Gene– 572 C/G Polymorphism and The

Risk of Type 2 Diabetes Mellitus: A Meta-Analysis Of 11,681 Subjects. *Annals of Human Genetics*, 114-106,77

Zhuanping, Z., Rifang, L., Qing, C. & Sidong, C. 2016. Association Between Interleukin 6, 10 Gene Polymorphisms and Risk of Type 2 Diabetes Mellitus in A Chinese Population. *Meta Gene*, 10, 13-17.