



Development and validation of a new analytical RP-HPLC method for simultaneous determination of Glibenclamide and Atenolol in bulk

Anitha P^{*1}, Ramkanth S², Satyanarayana S V³

¹Research Scholar, JNTUA, Anantapuramu, Andhra Pradesh, India

²Department of Pharmaceutics, Annamacharya College of Pharmacy, New Boyanapalli, Rajampet, Kadapa, Andhra Pradesh, India

³Department of Chemical Engineering, JNTUA, Anantapuramu, Andhra Pradesh, India

Article History:

Received on: 25.05.2019

Revised on: 12.08.2019

Accepted on: 19.08.2019

Keywords:

Atenolol,
Glibenclamide,
Combination,
HPLC,
Validation

ABSTRACT

A new, simple, reliable, fast, sensitive and economical RP-HPLC method was developed and validated for simultaneous estimation of two fixed-dose combinations frequently prescribed in coexisted chronic diseases such as diabetes (GLB) and hypertension (ATN) in bulk for the first time. The mobile phase used for the chromatographic runs consisted of 0.01N potassium dihydrogen ortho phosphate (pH 4.8) and acetonitrile (55:45, v/v). The separation was achieved on column (BDS C18 250 x 2.1mm, 1.6m) using isocratic mode. Drug peaks were well separated and were detected by a UV detector at 235.0 nm. The method was linear at the concentration range 2.5-15 µg/ml for Glibenclamide (GLB) and 6.25-37.5 µg/ml for Atenolol (ATN), respectively. The method has been validated according to ICH guidelines with respect to system suitability, specificity, precision, accuracy and robustness. The method was validated for system suitability, linearity, accuracy, precision, detection, quantification limits and robustness and was found it is acceptable in the range of 2.5-15 µg/ml for GLB and 6.25-37.5 µg/ml for ATN. The LOD and LOQ of GLB was found to be 0.48 µg/ml and 1.47 µg/ml and for ATN was found to be 0.72 µg/ml and 2.20 µg/ml, respectively. The method was applied to drug interaction studies of GLB with ATN to illustrate the scope and application of the methods to manage two different therapeutic classes of drugs, as they may co-administered in concurrent diseases.



*Corresponding Author

Name: Anitha P

Phone: 91-9908344380

Email: posina.anitha26@gmail.com

ISSN: 0975-7538

DOI: <https://doi.org/10.26452/ijrps.v10i3.1491>

Production and Hosted by

IJRPS | <https://ijrps.com>

© 2019 | All rights reserved.

INTRODUCTION

Diabetes mellitus (DM) is by defects in insulin activity, insulin secretion or both which is a metabolic

disorder characterized by hyperglycemia ([International Diabetes Federation, 2015](#); [Boyle et al., 2010](#)). The reason for the disease may be not enough insulin is produced in Type I DM (T1DM), glucose is not moved out into cells Type 2 DM (T2DM) ([Bilous and Donnelly, 2010](#)) and another one is gestational diabetes that may occur during pregnancy. Complications such as stroke, coronary heart diseases, nephropathy, neuropathy and retinopathy make great contributions to mortality ([Roglic and Unwin, 2010](#)).

Diabetes mellitus have two to fourfold higher death rate due to cardiovascular diseases than others ([Sowers et al., 2001](#)). Various treatment approaches for hypertension to reduce the risk of such complications ([Mogensen et al., 1991](#); [Fuller](#)

et al., 1983) in patients with T2DM. Stringent control over blood pressure, below those for the non-diabetic patient, has been shown to be effective, especially for a diabetic patient to lower the cardiovascular events (Aldington, 1998).

To achieve these aspirations in the majority cases, two to three drugs from different categories need to be prescribed (Munger, 2010). The chief oral antidiabetic drugs for diabetes mellitus are Metformin alone or in combination with the second-generation sulfonylureas viz. Glibenclamide (GLB), Gliclazide, Glimepiride, and Glipizide (Haupt *et al.*, 1991). As compared to other sulfonylurea's GLB provides convenient, effective and better tolerance (Giachetti *et al.*, 1997).

Beta-blockers /calcium channel blocker with a renin inhibitor (Bell, 2009) are prime agents for control of hypertension. When compared with the individual use, the combined form provides better control of hyperglycemia, insulin resistance and blood pressure. However, when they are combined the *in vivo* safety of the drugs may get altered. Therefore, the potential drug interactions of the combination of the Atenolol (ATN) and GLB in terms of pharmacokinetic perspective need to be studied before going further. There are numerous validated methods available for analysis of ATN (Davidson *et al.*, 2004; Aburuz *et al.*, 2005; Georgita *et al.*, 2007) and GLB (Mistri *et al.*, 2007; Johnson and Lewis, 2006) alone. There was no reported validated method for their simultaneous quantization in any type of biological sample.

The concept of combining agents with manifestly different indications has become a certainty of treatment in patients with a chronic disease where it requires treatment for several different pharmacotherapies (Porwal and Talele, 2017). As a primary treatment to diabetic hypertensive subjects, generally, a fixed-dose of GLB and ATN or combination of drugs is commonly prescribed (Sengupta *et al.*, 2017). Fixed-dose combination of GLB and ATN shows effective control on glucose balance and better BP control in a diabetic hypertensive patient which are not tested literally. Given this background, the present study was envisaged to develop and validate the HPLC method as per ICH guidelines. There is no analytical method available to simultaneously quantitate ATN and GLB in a single run till date.

EXPERIMENTAL PART

Chemicals and reagents

GLB obtained from Sri Raghavendra Chemicals and

Suppliers, Bangalore. ATN obtained from Indian Drugs, Hyderabad. Analytical/HPLC grade chemicals and solvents were obtained from Rankem (Hyderabad, India).

Chromatographic apparatus and Condition

A high-performance liquid chromatographic system (Water 2690) composed of PDA-2996 detector. Chromatographic integration, data acquisition and recording were performed by Empower 2 software. Optimization of the chromatographic condition is by using different mobile phase combination, columns and organic phases. The mobile phase flows with a flow rate of 1.0 ml /min. For retention of GLB & ATN (Figure 1) and a combination of actives, various HPLC methods were optimized and validated. Optimum separation conditions were obtained with BDS C18 250 x 2.1mm, 1.6m column, mobile phase taken in the ratio 55:45 consisting of 0.01N potassium dihydrogen ortho phosphate (pH 4.8) and acetonitrile (ACN) in gradient mode with an injection volume 1.0 ml with column oven temperature maintained at 30°C and elution monitored by a detector wavelength at 235.0 nm. Various methods have been described for the individual drug determination (El-Saharty, 2003; Sultana *et al.*, 2008), but for the simultaneous analysis of these drugs, no method is reported till date.

Preparation of standard solution

2.5 mg of GLB & 6.25 mg of ATN were accurately weighed and diluted to 25 ml and add 10 ml of diluents. After 10min of sonication, the solution is diluted with diluents (100 µg/ml GLB & 250 µg/ml ATN). 1ml of the standard solution was diluted to 10ml with diluents (10µg/ml GLB & 25µg/ml ATN).

Method validation

According to ICH Guidelines, for ATN and GLB assay, the RP-HPLC method was validated by linearity, accuracy, precision, LOD, LOQ, and robustness.

Accuracy

Nearness between the expected value and the obtain value (value found) gives the analytical method accuracy. Accuracy of the developed method is by successive analysis (n = 3) of three different concentrations (50%, 100% and 150%) of standard drugs solution. Percent recovery (R %) of an analyte recovered is obtained. The data of the results were statistically analyzed.

Precision

When the method is applied repetitively in three different occasion's precision of a technique is the degree of agreement among individual tests. By analyzing the calibration curves of different concentra-

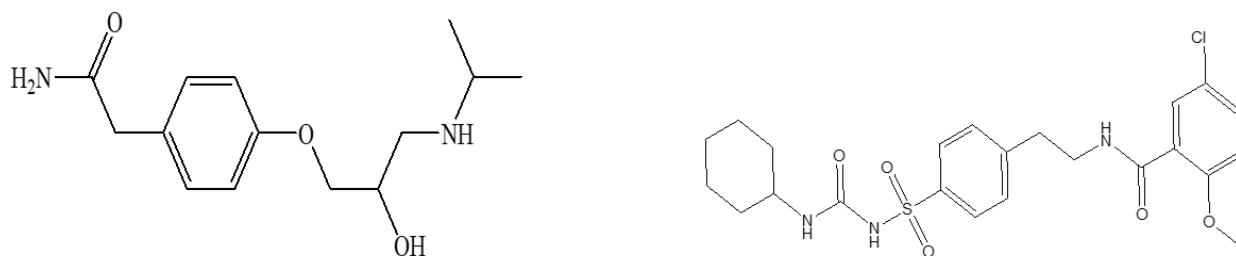


Figure 1: Chemical structure of ATN and GLB

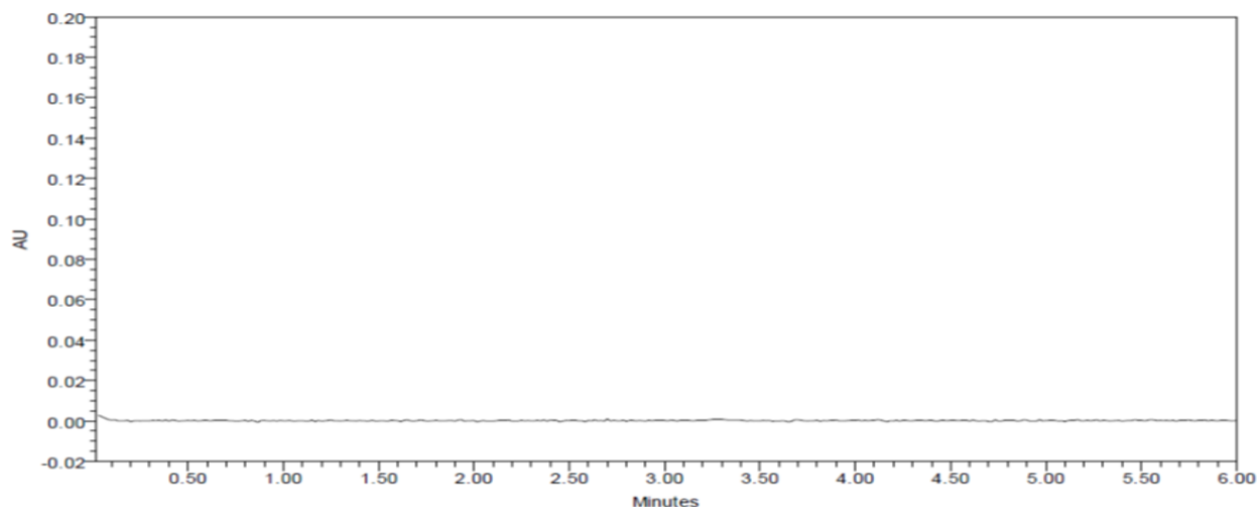


Figure 2: HPLC chromatograms of blank mobile phase spike (chromatogram A) and HPLC chromatograms of a mobile phase containing 10 $\mu\text{g/ml}$ GLB & 25 $\mu\text{g/ml}$ ATN (chromatogram B)

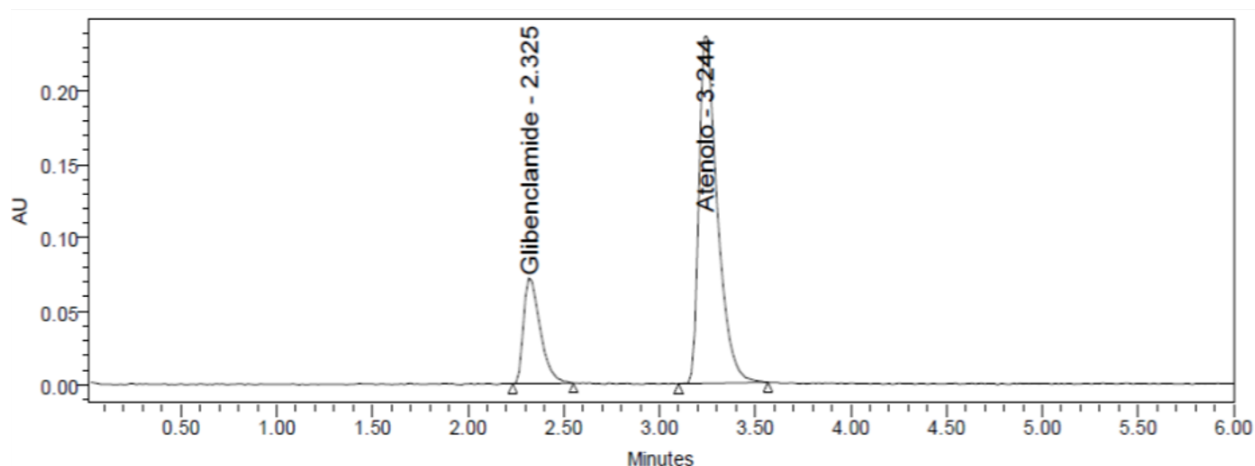


Figure 3: HPLC chromatograms of mobile phase containing 10 $\mu\text{g/ml}$ GLB & 25 $\mu\text{g/ml}$ ATN (chromatogram B)

tions of drugs on three different days and within the same day respectively in six replicates, the inter-day precision and intra-day precision was determined. The relative standard deviation (%RSD) of the method was expressed as total precision, where % RSD \leq 2% was accepted.

Linearity

Linearity is the ability to obtain results of the experiment is directly proportional to the analyte concentration. Linearity was determined by a minimum

of six calibration levels (25, 50, 75, 100, 125 and 150%). Coefficient of correlation, slope and intercept were used to estimate the linearity by using the calibration curve. The correlation coefficient (r^2) > 0.998 value is accepted for the data to the regression line.

LOD and LOQ

Limit of detection (LOD) and limit of quantification (LOQ) were considered based on the signal-to-noise ratio, which is 3 & 10, respectively. Actually, it is

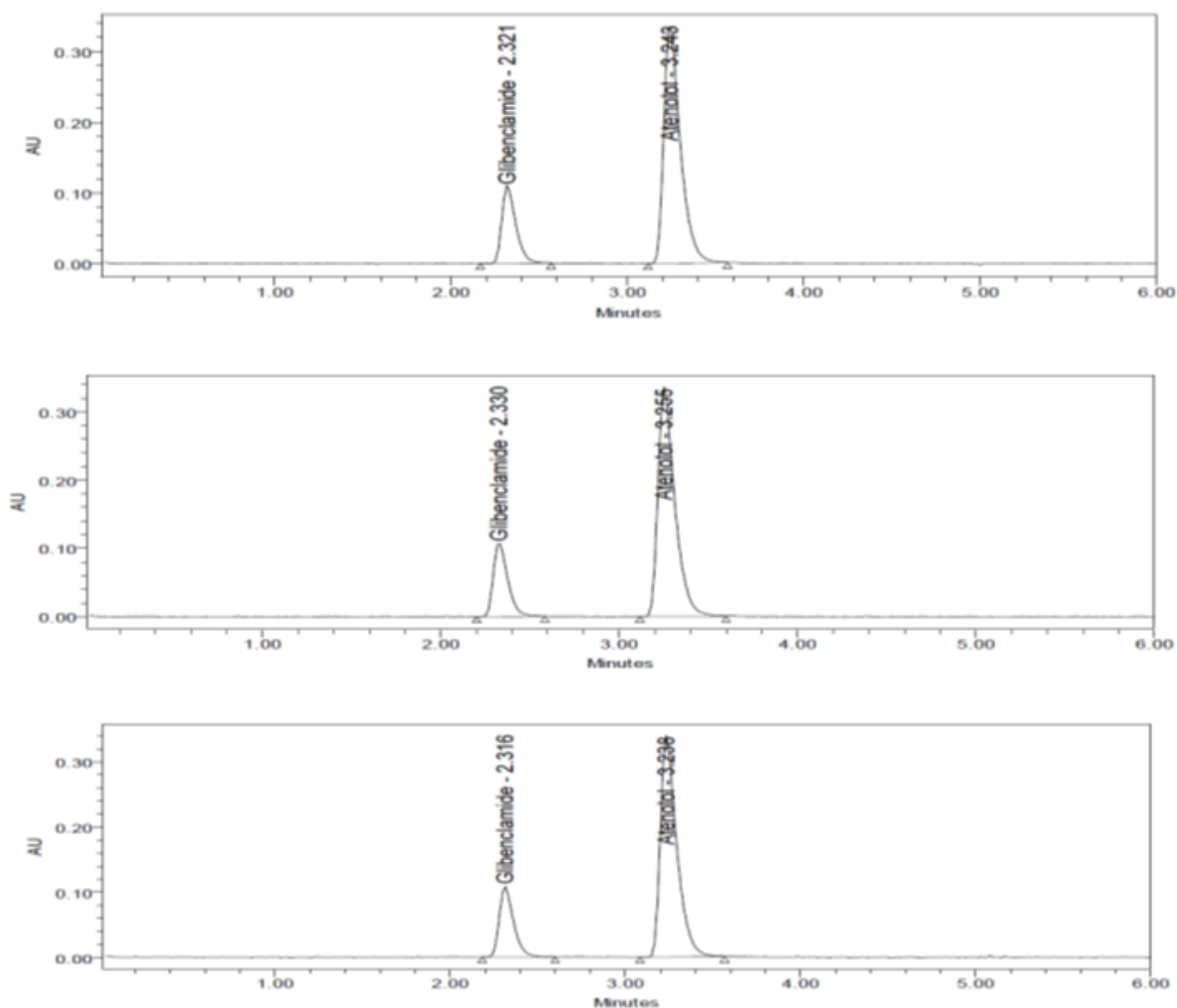


Figure 4: Chromatograms at 50% Recovery level (Triplicate)

established by the minimum concentration at which the analyte can be reliably detected and quantified by comparing blank samples with a known low concentration of an analyte.

Robustness

For robustness, the chromatographic conditions were changed deliberately using HPLC method. Reliability during normal usage is measured by its capacity to remain unaffected by small, but deliberate variations in method parameters.

Degradation studies

Stress factors suggested for common forced degradation studies such as acid degradation, base degradation, oxidation, photolysis, thermal degradation (FDA, 2000; Blessy *et al.*, 2014).

Sample Preparation

10mg of GLB and 25mg of ATN powder were accurately weighed and diluted to 50 ml, by adding diluents and sonicated for 25 min, and filtered by HPLC

filters (200 μ g/ml GLB & 500 μ g/ml ATN). Finally, filtered sample solution of 0.5ml was diluted to 10ml with diluents (10 μ g/ml GLB & 25 μ g/ml ATN).

Acid Degradation Studies

1ml of 5N Hydrochloric acid was added to 1 ml of sample solution GLB & ATN, and at 60°C it is refluxed for 30mins. The final solution was diluted to obtain a 10ppm, & 25ppm solution and injected (10 μ L) solution into the system. The stability of the sample solution was determined by analysing chromatograms (Singh and Bakshi, 2000).

Alkali Degradation Studies

5N Sodium hydroxide was added to 1 ml of sample solution GLB & ATN, and at 60°C it is refluxed for 30 min. 10 μ L of resulting solution (10ppm & 25ppm) was injected into the system and evaluate the stability of the sample by recorded chromatograms (Alsante *et al.*, 2007).

Peroxide degradation

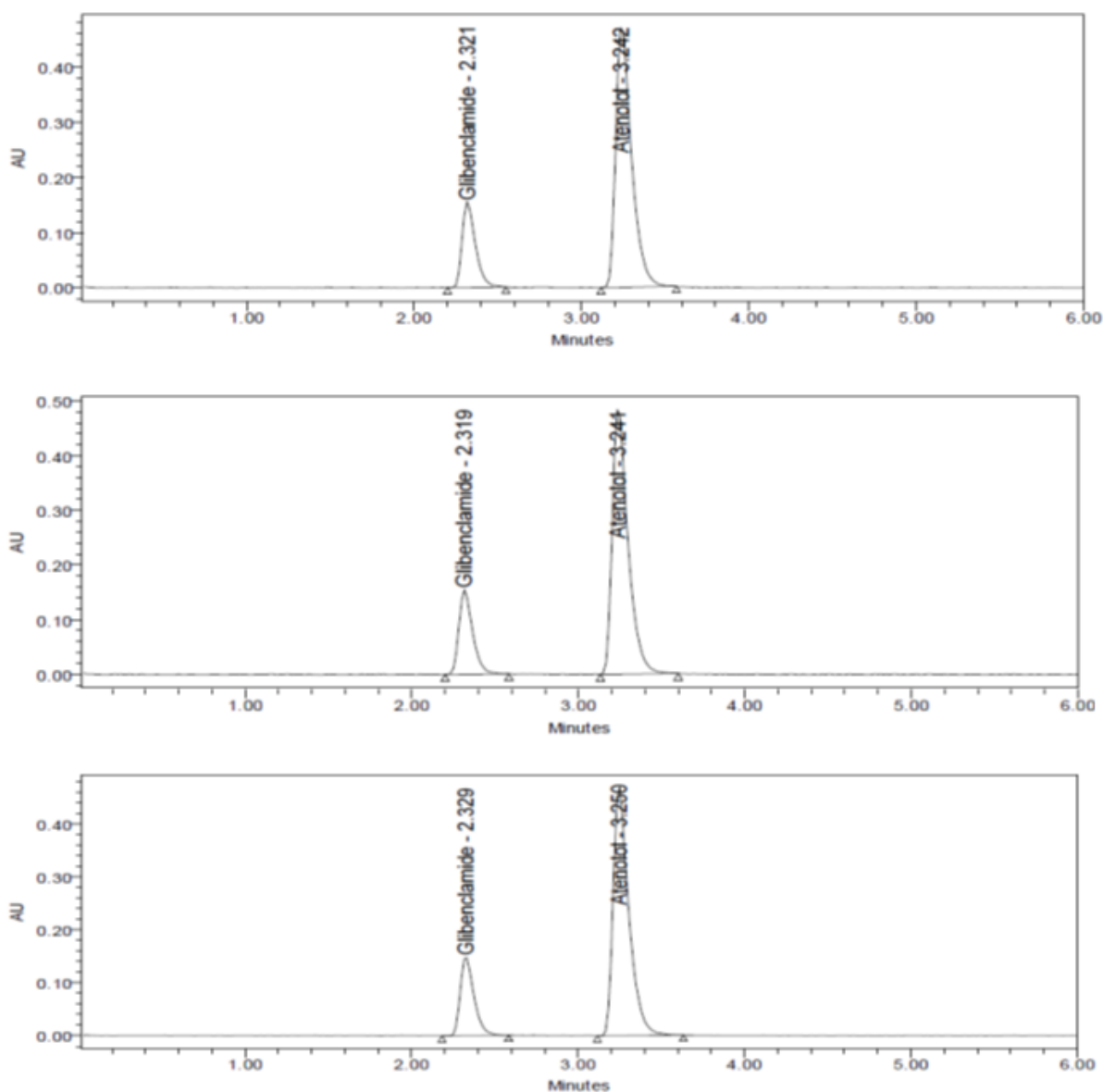


Figure 5: Chromatograms at 100% Recovery level (Triplicate)

Table 1: System suitability Results for HPLC

System suitability Parameters	GLB	ATN
Retention time	2.339	3.302
Area	408922	1041734
USP Plate Count	3595.0	5584.1
USP Tailing	1.5	1.5
Resolution	-	5.6

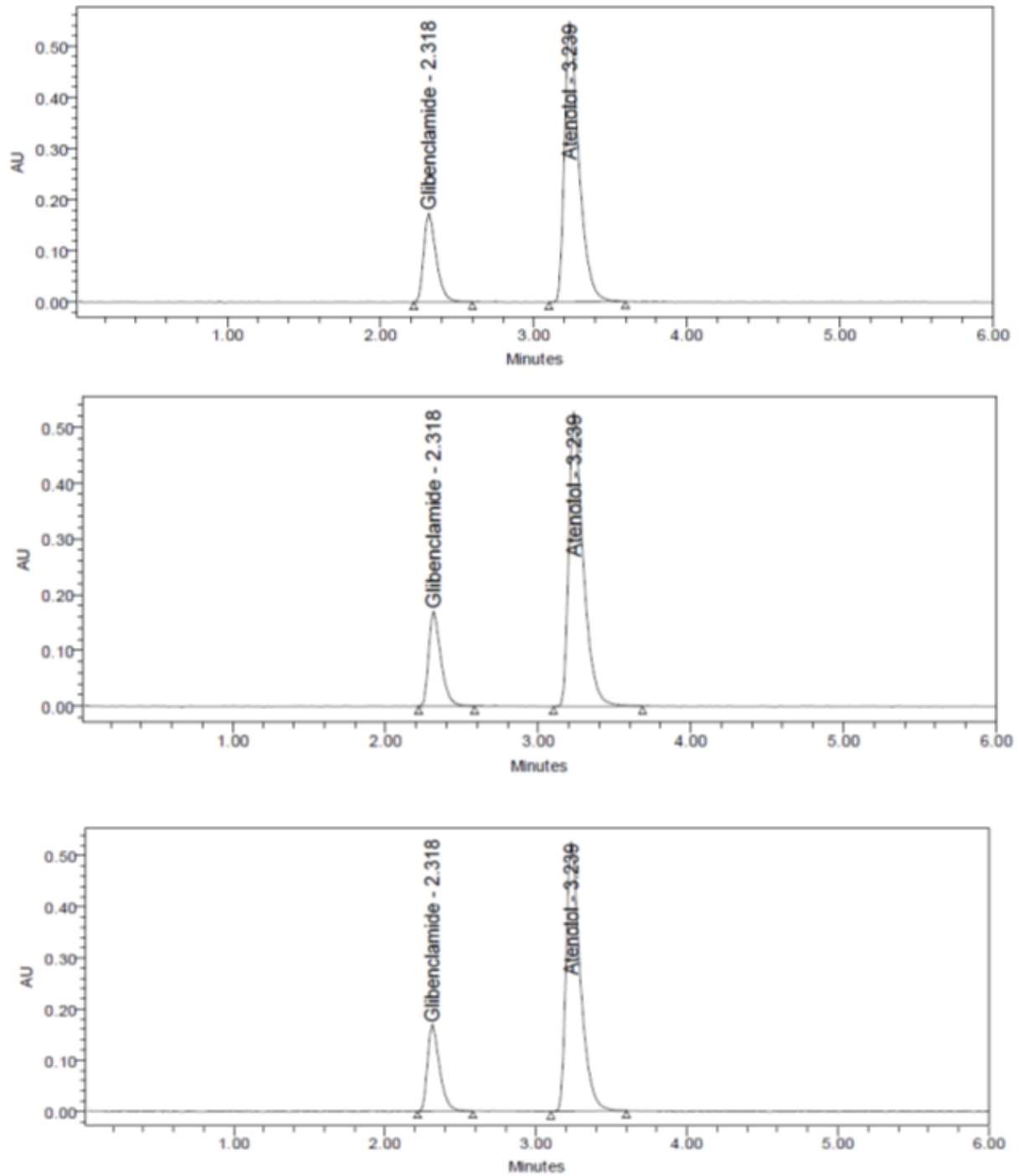


Figure 6: Chromatograms at 150% Recovery level (Triplicate)

Table 2: Recovery studies of GLB and ATN

Drugs	% Level of recovery	Mean %Recovery
GLB	50	99.15±0.12
	100	99.94±0.87
	150	99.77±0.44
ATN	50	99.24±0.88
	100	100.10±0.69
	150	99.97±0.76

Table 3: System Precision studies of GLB and ATN

S.No	Precision	GLB		ATN	
		Mean ± SD	% RSD	Mean ± SD	% RSD
1	Repeatability	406510 3604.4	± 0.9	1067155±4525.9	0.4
2	Intermediate Precision	394228 3959.0	± 1.0	984912±4210.8	0.4

Table 4: Linearity studies of GLB and ATN

S. No	Concentration of GLB (ppm)	AUC for GLB	Concentration of ATN (ppm)	AUC for ATN
1	0	0	0	0
2	2.5	107857.6	6.25	256152.3
3	5	204184.6667	12.5	506781
4	7.5	315382	18.75	754455
5	10	404592.6667	25	1040950
6	12.5	515066.3333	31.25	1265461
7	15	611057.3333	37.5	1503674
8	Slope	40685.7	Slope	40364.7
9	Y-Intercept	3162.66	Y-Intercept	4230.7
10	R2	0.999	R2	0.999

Table 5: Robustness results for both GLB and ATN (acceptance limit RSD% < 2)

S.No	Robustness Parameter	% RSD for GLB	% RSD for ATN
1	Flow rate (0.9 ml/min)	0.4	0.91
2	Flow rate (1.1ml/min)	0.9	0.6
3	Temperature 25°C	0.4	0.4
4	Temperature 35°C	0.79	0.7
5	Mobile phase (50B:50 A)	0.73	0.7
6	Mobile phase (60B:40A)	0.77	1.0

Table 6: Forced Degradation Studies of GLB and ATN

Stress Condition (Degradation)	GLB			ATN		
	Relative Amount (%)	% Degradation	Assay (%)	Relative Amount (%)	% Degradation	Assay (%)
Acid	94.61	5.39	94.61	92.21	7.79	92.21
Alkali	95.36	4.64	95.36	95.66	4.34	95.66
Peroxide	91.32	8.68	91.32	93.18	6.82	93.18
Thermal	97.30	2.70	97.30	97.67	2.33	97.67
UV	98.33	1.67	98.33	98.30	1.70	98.30
Water	98.66	1.34	98.66	99.44	0.56	99.44

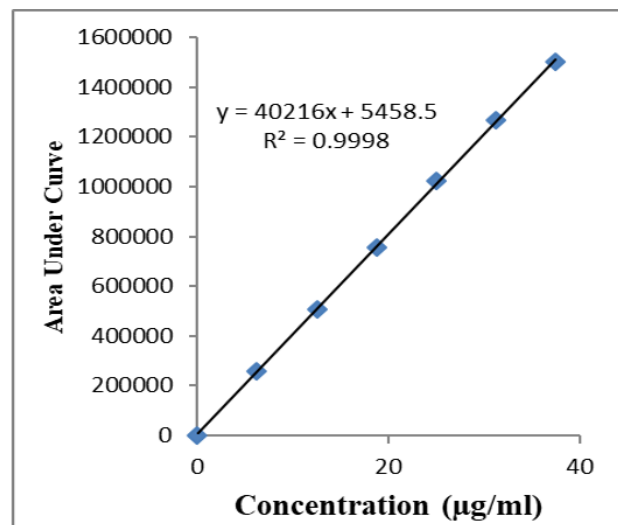
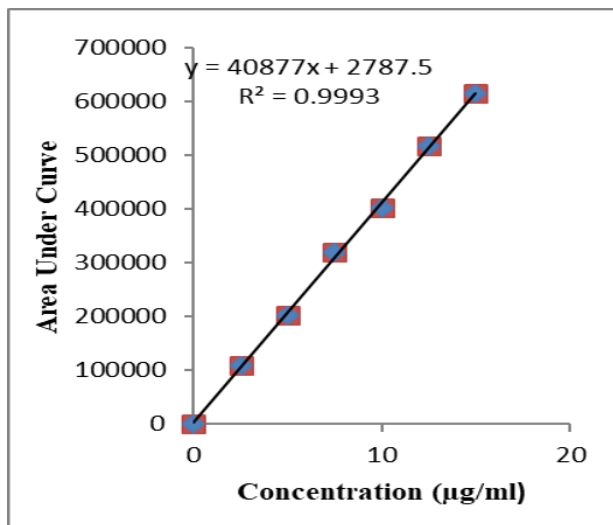


Figure 7: Linearity curves for GLB and ATN

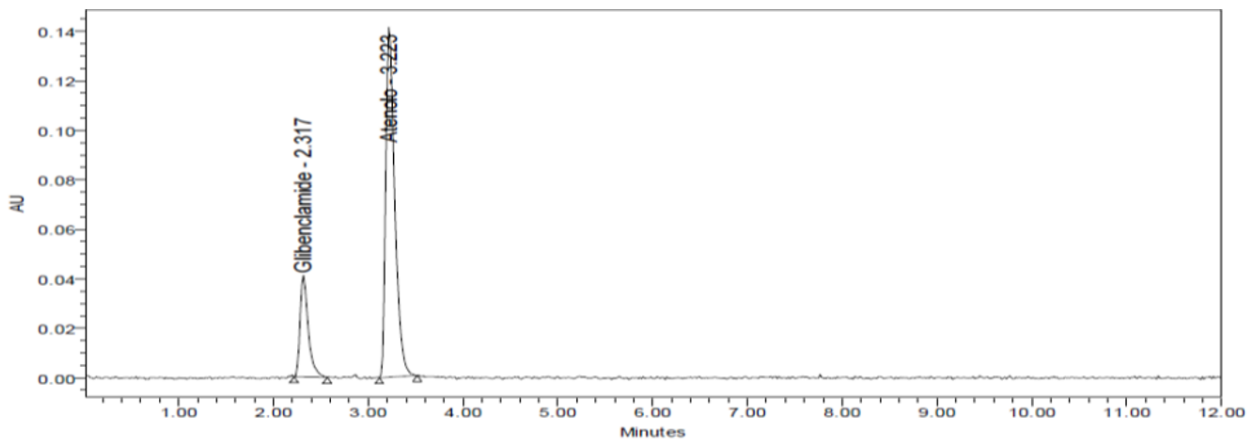


Figure 8: Chromatogram for Acid degradation

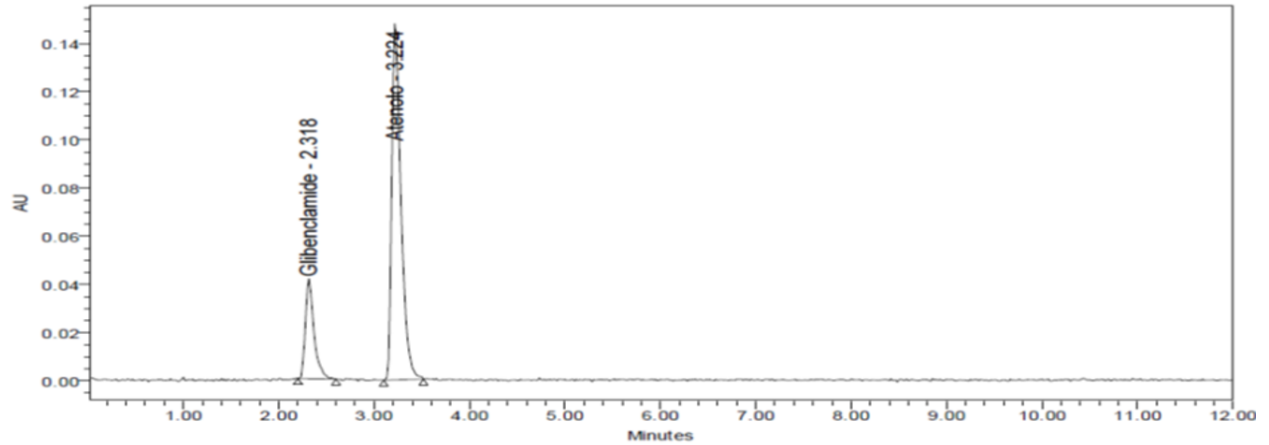


Figure 9: Chromatogram for Base degradation

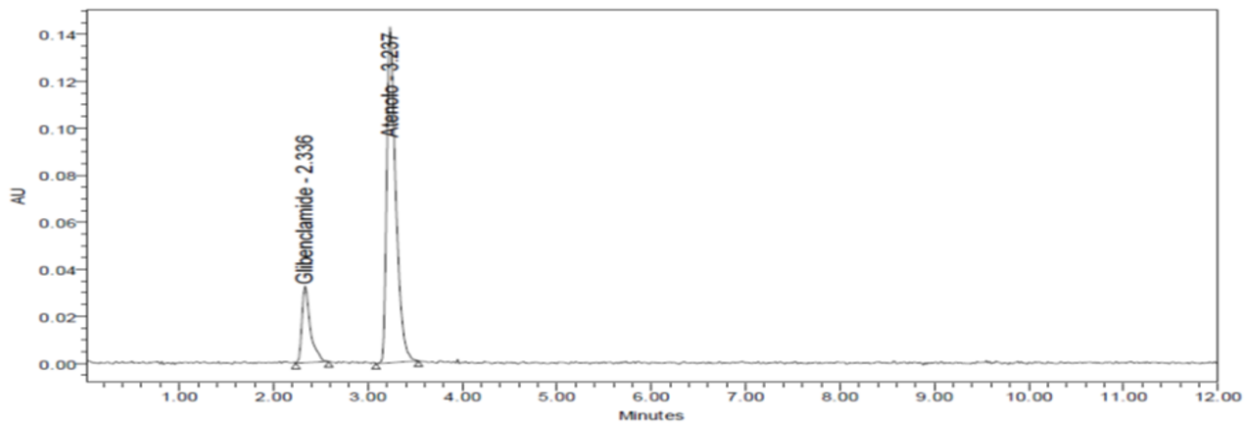


Figure 10: Chromatogram for Peroxide degradation

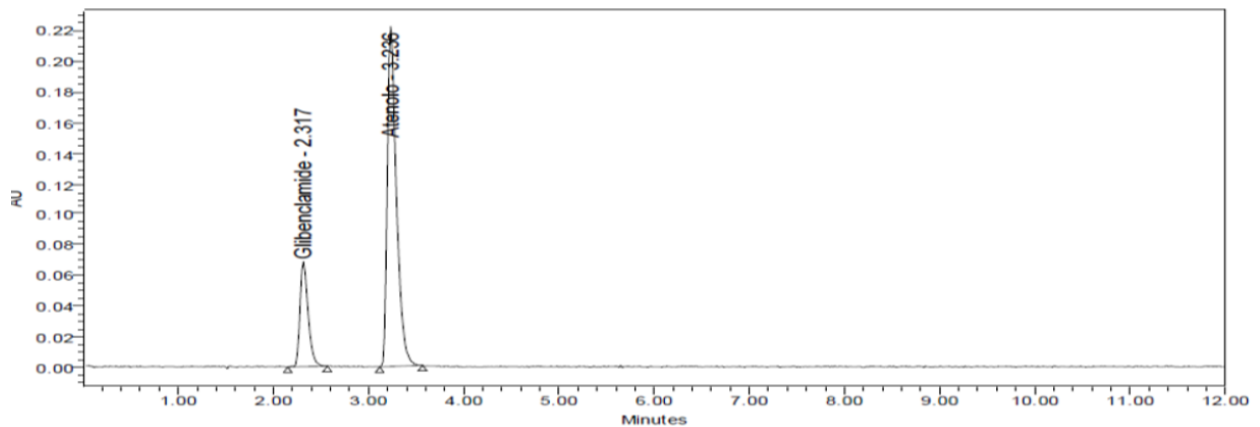


Figure 11: Chromatogram for Thermal degradation

30% hydrogen peroxide (H_2O_2) was added to 1 ml of sample solution of GLB & ATN, which is refluxed for 30 min at $60^\circ C$. The final solution (10 ppm & 25 ppm) which was obtained by dilution, $10 \mu L$ of it injected into the system for assessing the stability of sample (Boccardi, 2005).

Thermal Degradation Studies

For Thermal degradation, the Sample solution was kept in an oven at $105^\circ C$ for 6 h. For assess-

ing the stability of the sample by recording the chromatograms, the resultant solution which was diluted to (10 ppm & 25 ppm) solution, $10 \mu L$ of it was injected into the system (Qiu and Norwood, 2007).

Photo Stability studies

The sample solution was kept for 7 days in UV Light chamber or in photo stability chamber at 200 Watt h/m^2 for studying photo stability studies. For

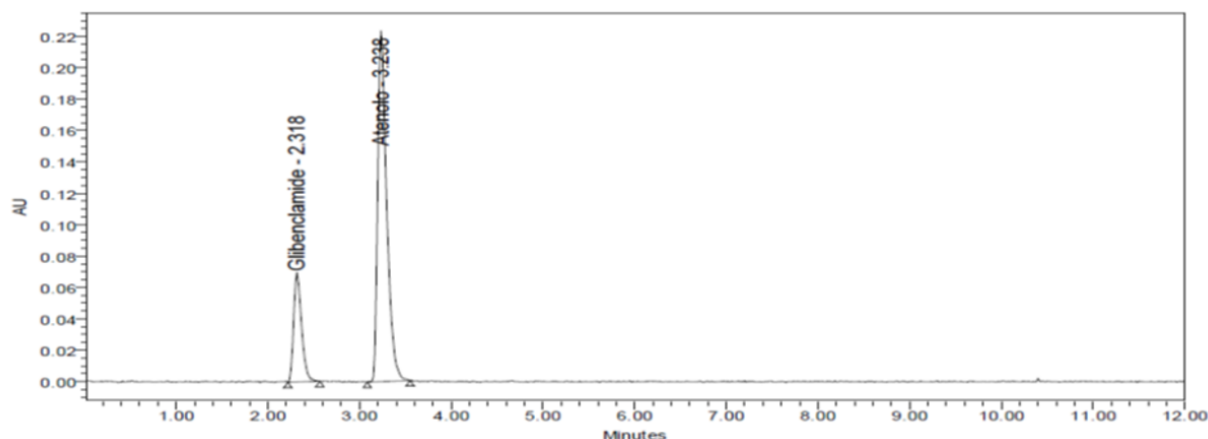


Figure 12: Chromatogram for UV degradation

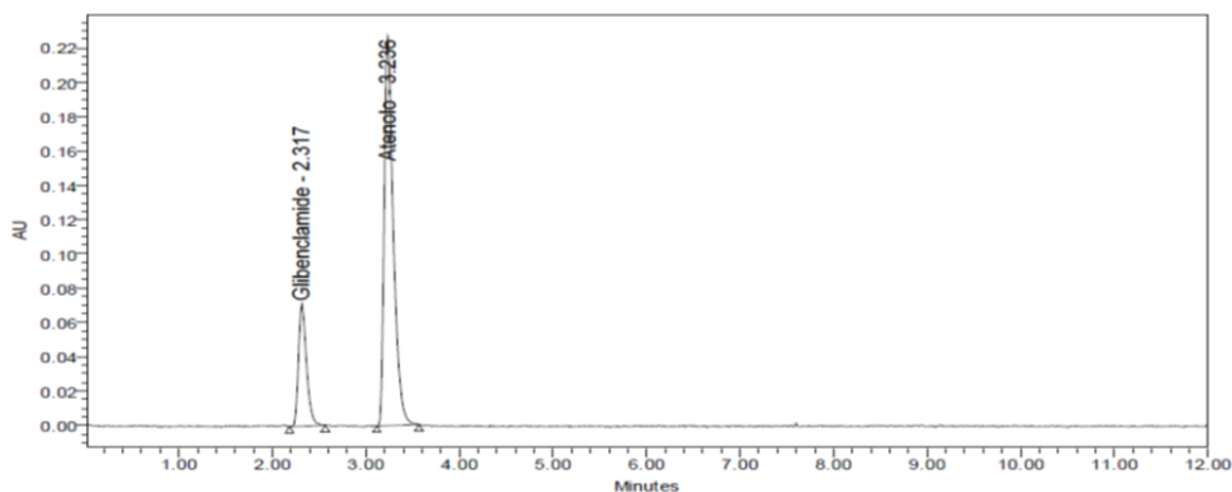


Figure 13: Chromatogram for Water degradation

assessing the stability of the sample solution, the reult diluted solution (10 ppm & 25 ppm) of 10 μ L were injected into the system (Ahuja and Scypinski, 2001).

Neutral Degradation Studies

Neutral degradation studies were studied y drug refluxing in water for 6hrs at a tem erature of 60 °C. The resultant diluted solution (10 ppm & 25ppm), 10 μ L of it were i jected into the system for assessment of the stability of the sample (Blessy et al., 2014).

RESULTS AND DISCUSSION

Development and optimization of HPLC method

An isocratic LC method, coupled with PDA detection, was developed for the simultaneous determination of ATN and GLB. Chromatogram A and chromatogram B represents the blank mobile phase and an average retention time of 2.322 min for GLB and 3.260 min for ATN, and with no interfering peaks respectively (Figure 2&Figure 3), the suitability

parameters were given Table 1, which indicates the specificity of the method. By using BDS C18 250 x 2.1mm, 1.6m column and gradient elution technique using a mobile phase composed of Buffer: ACN pH 4.8 in a ratio of 55:45 and the most suitable separation with the highest resolution between peaks of GLB and ATN was achieved. However, drugs peaks are obtained at shorter retention times at a temperature of 30°C compared to those at 25°C. So, 30°C was chosen for the chromatographic measurement. According to the buffer capacity of phosphate, the pH of the buffer was selected, which leads to less variation in retention time and also resisted to pH changes.

According to ICH guidelines (International Council for Harmonisation), this method was validated. The following validation characteristics were addressed.

Accuracy

The developed HPLC method was assessed for accuracy in a laboratory at three different concentration levels (50%, 100% and 150%) within the working linearity range of both drugs of a standard mix-

ture of ATN and GLB. Accuracy was expressed as the recovery% (ICH, 2005) and standard error for GLB and ATN. Table 2 and Figures 4, 5 and 6 shows the method accuracy for both drugs with satisfactory recovery%. These results reveal that a developed method was found to be accurate.

Precision

Information on the random errors was given by the precision of a given method. The nearness of values obtained from multiple sampling of the same homogenous sample under prescribed condition (ICH, 2005) between a series of measurements. Both repeatability (intra-day precision) and intermediate precision (inter-day precision) were obtained. The results for precision were found to be within permissible limits (1% & 2% respectively) Table 3.

Repeatability (intra-day)

The Repeatability of the method was obtained by using the same analytical procedure, within the same laboratory on the same day, using the same equipment, by the same analyst between a series of measurements. Repeatability is by RSD% values obtained by repeating the assay (Table 3).

Intermediate precision (Inter-day)

The Intermediate precision was carried out as described under repeatability but on three successive days, each of two determinations (n =6) (Table 3).

The standard for intra-day and inter-day precisions requires a RSD less than 2%.

Linearity and Range

The linearity of the method was assessed at a range of 2.5–15 ppm GLB and 6.25–37.5 ppm ATN. Calibration curves were plotted by taking peak areas against concentrations (Table 4 and Figure 7).

LOD and LOQ

LOD is the minimum concentration which can be detected, not quantified and LOQ is the minimum concentration with satisfactory precision and accuracy in a ratio of 3:1 and 10:1 respectively. The LOD and LOQ of GLB was found to be 0.48 $\mu\text{g/ml}$ and 1.47 $\mu\text{g/ml}$ and for ATN was found to be 0.72 $\mu\text{g/ml}$ and 2.20 $\mu\text{g/ml}$ respectively.

Robustness

One can determine the considerable or significant influence of factors on analytical results by performing a set of experiments and combining changes in conditions (see Table 5). No significant effect in resolution between two drugs peaks when there

are small variations in column temperature, chromatographic flow and mobile phase composition (% acetonitrile) which was confirmed by the Standard means and averages.

Degradation Studies

Degradation Studies was carried out to exhibit explicitly the method developed to measure the changes in concentration of two drugs. For validation of chromatographic assays, between 5% and 20% has been accepted (Szepesi *et al.*, 1991; Carr and Wahlich, 1990). For analytical validation, 10% degradation is optimal (Jenke, 1996; Blessy *et al.*, 2014). The chromatograms of two drugs degradation are shown in Figures 8, 9, 10, 11, 12 and 13. For quantitative analysis of the drugs under various stress, the condition was done by the proposed HPLC method. The contents of stress testing are shown in Table 6. Under our developed auto-injector device, the combination drugs solution was stable and within the acceptance criteria in all degradation condition.

CONCLUSION

For simultaneous determination of ATN and GLB the developed method is suitable and valid because of small retention time separation for both peaks and good resolution. It can be used for routine analysis of GLB and ATN in combination products which reduces the multiple medical regimens. This method is simple, specific, linear, accurate and precise. Has ability to separate the two drugs which are usually found in the serum of diabetic hypertensive patients. The results of the present study benefits in the studies of drug interaction and also very profitable for pharmaceutical companies.

REFERENCES

- Aburuz, S., Millership, J., Mcelnay, J. 2005. The development and validation of liquid chromatography method for the simultaneous determination of metformin and glipizide, gliclazide, glibenclamide or glimiperide in plasma. *Journal of Chromatography B*, 817(2):277–286.
- Ahuja, S., Scypinski, S. 2001. Handbook of Modern Pharmaceutical Analysis. 1:23. ISBN: 9780080488929.
- Aldington, S. 1998. Tight blood pressure control and risk of macrovascular and microvascular complications in type 2 diabetes (UKPDS 38) UK Prospective Diabetes Study Group (UKPDS). *BMJ*, 317:703–713. BMJ.
- Alsante, K., Ando, A., Brown, R., Ensing, J., Hatajik,

- T., Kong, W., Tsuda, Y. 2007. The role of degradant profiling in active pharmaceutical ingredients and drug products☆. *Advanced Drug Delivery Reviews*, 59(1):29–37.
- Bell, D. S. H. 2009. Treatment of diabetic hypertension. *Diabetes, Obesity and Metabolism*, 11(5):433–444.
- Bilous, R., Donnelly, R. 2010. Handbook of Diabetes. 4. ISBN:9781444391374.
- Blessy, M., Patel, R. D., Prajapati, P. N., Agrawal, Y. K. 2014. Development of forced degradation and stability indicating studies of drugs-A review. *Journal of Pharmaceutical Analysis*, 4(3):159–165.
- Boccardi, G. 2005. Pharmaceutical Stress Testing. pages 220–624. Oxidative susceptibility testing.
- Boyle, J. P., Thompson, T. J., Gregg, E. W., Barker, L. E., Williamson, D. F. 2010. Projection of the year 2050 burden of diabetes in the US adult population: dynamic modeling of incidence, mortality, and prediabetes prevalence. *Population Health Metrics*, 8(1):29–29.
- Carr, G. P., Wahlich, J. C. 1990. A practical approach to method validation in pharmaceutical analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 8(8-12):613–618.
- Davidson, J. A., Scheen, A. J., Howlett, H. C. S. 2004. Tolerability Profile of Metformin/Glibenclamide Combination Tablets (Glucovance®). *Drug Safety*, 27(15):1205–1216.
- El-Saharty, Y. 2003. Simultaneous high-performance liquid chromatographic assay of furosemide and propranolol HCL and its application in a pharmacokinetic study. *Journal of Pharmaceutical and Biomedical Analysis*, 33(4):699–709.
- FDA 2000. Guidance for industry. Analytical procedures and methods validation. Chemistry, manufacturing, and controls documentation. *Draft Guidance*. Draft Guidance.
- Fuller, J. H., Shipley, M. J., Rose, G., Jarrett, R. J., Keen, H. 1983. Mortality from coronary heart disease and stroke in relation to degree of glycaemia: the Whitehall study. *British Medical Journal*, (6396):867–870. Clinical Research Ed.
- Georgita, C., Albu, F., David, V., Medvedovici, A. 2007. Simultaneous assay of metformin and glibenclamide in human plasma based on extraction-less sample preparation procedure and LC/(APCI)MS. *Journal of Chromatography B*, 854(1-2):211–218.
- Giachetti, C., Tenconi, A., Canali, S., Zanolio, G. 1997. Simultaneous determination of atenolol and chlorthalidone in plasma by high-performance liquid chromatography application to pharmacokinetic studies in man. *Journal of Chromatography B: Biomedical Sciences and Applications*, 698(1-2):187–194.
- Haupt, E., Knick, B., Koschinsky, T., Liebermeister, H., Schneider, J., Hirche, H. 1991. Oral antidiabetic combination therapy with sulphonylureas and metformin. *Diabete & metabolisme*, 17(1):224–231. Pt 2.
- International Diabetes Federation 2015. IDF Diabetes Atlas. 7:1–23.
- Jenke, D. R. 1996. Chromatographic Method Validation: A Review of Current Practices and Procedures. I. General Concepts and Guidelines. *Journal of Liquid Chromatography & Related Technologies*, 19(5):719–736.
- Johnson, R. D., Lewis, R. J. 2006. Quantitation of atenolol, metoprolol, and propranolol in postmortem human fluid and tissue specimens via LC/APCI-MS. *Forensic Science International*, 156(2-3):106–117.
- Mistri, H. N., Jangid, A. G., Shrivastav, P. S. 2007. Liquid chromatography tandem mass spectrometry method for simultaneous determination of antidiabetic drugs metformin and glyburide in human plasma. *Journal of Pharmaceutical and Biomedical Analysis*, 45(1):97–106.
- Mogensen, C. E., Hansen, K. W., Pedersen, M. M., Christensen, C. K. 1991. Renal Factors Influencing Blood Pressure Threshold and Choice of Treatment for Hypertension in IDDM. *Diabetes Care*, 14:13–26. Supplement 4.
- Munger, M. A. 2010. Polypharmacy and Combination Therapy in the Management of Hypertension in Elderly Patients with Co-Morbid Diabetes Mellitus. *Drugs & Aging*, 27(11):871–883.
- Porwal, P. K., Talele, G. S. 2017. Development of validated HPLC-UV method for simultaneous determination of Metformin, Amlodipine, Glibenclamide and Atorvastatin in human plasma and application to protein binding studies. *Bulletin of Faculty of Pharmacy*, 55(1):129–139.
- Qiu, F., Norwood, D. L. 2007. Identification of Pharmaceutical Impurities. *Journal of Liquid Chromatography & Related Technologies*, 30(5-7):877–935.
- Roglic, G., Unwin, N. 2010. Mortality attributable to diabetes: Estimates for the year 2010. *Diabetes Research and Clinical Practice*, 87(1):15–19.
- Sengupta, P., Chatterjee, B., Pal, T. K. 2017. Assessment of preclinical pharmacokinetics and acute toxicity of pioglitazone and telmisartan combination. *Regulatory Toxicology and Pharmacology*,

- 91:151–158.
- Singh, S., Bakshi, M. 2000. Guidance on Conduct of Stress Tests to Determine Inherent Stability of Drugs. *Pharmaceutical Technology On-Line*, 24:1–14.
- Sowers, J. R., Epstein, M., Frohlich, E. D. 2001. Diabetes, Hypertension, and Cardiovascular Disease. *Hypertension*, 37(4):1053–1059.
- Sultana, N., Arayne, M. S., Iftikhar, B. 2008. Simultaneous Determination of Atenolol, Rosuvastatin, Spironolactone, Glibenclamide and Naproxen Sodium in Pharmaceutical Formulations and Human Plasma by RP-HPLC. *Journal of the Chinese Chemical Society*, 55(5):1022–1029.
- Szepesi, G., Gazdag, M., Mihályfi, K. 1991. Selection of high-performance liquid chromatographic methods in pharmaceutical analysis. *Journal of Chromatography A*, 464:94245–94251.