

# INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACEUTICAL SCIENCES

Published by JK Welfare & Pharmascope Foundation

# AMPA receptor localization in trigeminal ganglion and its upregulation in migraine

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ISSN: 0975-7538

DOI: https://doi.org/10.26452/ijrps.v10i3.1453

Production and Hosted by

IJRPS | https://ijrps.com

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#### **INTRODUCTION**

Glutamate is an excitatory neurotransmitter in the neurons which are responsible for degeneration leading to pathogenesis. Glutamate acts on two kinds of receptors Ionotrophic and Metabotrophic, which have a different pathway of activation. Ionotropic receptors are N-Methyl D-Aspartate (NMDA) receptors,  $\alpha$ -Amino-3-hydroxy-5-Methyl-4-isoxazolePropionic Acid (AMPA) and Kainate

receptors, which acts through voltage-gated sodium and calcium channels (Bonsi et al., 2005). The metabotropic receptors are of three groups (I, II, III) which consist of mGluR1-8 acts by G proteincoupled receptor mechanism. Blocking these ionotrophic glutamate receptors in the peripheral tissues decreases pain behaviour and nociception in inflammatory models (Julio-Pieper et al., 2011).

Chronic migraine characterized by moderate to severe episodes of headache very much disabling the normal day to day activities and accounts for 20% of all neurology outpatient consultation (Stone et al., 2010). Migraine is one of the neurological conditions listed by the World Health Organization, with a prevalence of 6-8% in male and 15-25% in female (Pietrobon and Striessnig, 2003). The hypothesis of migraine involves both vascular and neuronal components triggering the migraine attacks through trigeminovasular system, which is present in the brain stem.

Studies have shown that glutamate expression increased in the dorsal root ganglion after periph-

eral nerve injury indicating glutamate involvement in nociception (Kung *et al.*, 2013; Lee and Ro, 2007). Based on those studies, the AMPA receptors acting by voltage-gated calcium channel could play a major role in pathological excitations leading to nociception. The AMPA [receptors are of 4](#page-3-4) [types GluR 1-4 ou](#page-3-5)t of which GluR1 is present in all neurons innervated by both myelinated and unmyelinated fibers (Lee *et al.*, 2002). The glutamate receptors from the cell body can be exported to the nerve endings in the skin, muscles and joints (Lee and Ro, 2007) and followed by any inflammation these receptors get [sen](#page-3-6)[sitized and](#page-3-6) increase its expression (Du *et al.*, 2006). This study was done to identify the GluR1 receptors in the cells of trigemina[l ganglion and its](#page-3-5) changes after nociceptive stimulation.

## **METHODOLOGY**

After approval from the Institute Animal Ethics Committee, the male Wistar rats were obtained from the experimental animal lab of Saveetha Medical College and Hospital, Chennai. The rats of 12 numbers weighing from 200g to 250g were used and kept in cages with not more than three in a single cage. They were maintained at 12hr: 12hr light/dark cycles with water and food available ad libitum.

#### **Experimental Design**

The rats were divided into three groups

**Group 1**: Control rats (n=6) – Saline treated rats

**Group 2**: Migraine model rats (n=6) – nitroglycerin (10mg/kg subcutaneous back of neck) induced rat migraine model for 7 days.

#### **Tissue collection**

After seven days, the rats of the experiment, the rats were sacrificed and fixed. The fixation was done thoroughly using 4% paraformaldehyde perfused through transcardiac approach for one hour. Dissection was done to open the skull and brain was lifted to identify trigeminal ganglion. The trigeminal ganglion of all groups were collected separately and labeled then kept for overnight fixation. The tissues were sectioned (40*µ* thick) using optimum cutting temperature medium (OCT medium) with a cryostat and collected in multivial culture plates separately for each group.

#### **Immunohistochemistry**

The antibodies for GluR1 subunits of AMPA receptors were obtained from sigma laboratories, and the standard dilution ratio (1:250) was determined after repeated histochemical localization at various dilution ratios. The free-floating sections of trigem-

inal ganglion were localized for GluR1 subunits, focused by JENOPTIK ProgRes Capture Pro 2.7 (Germany) captured using ProgRes image capture software.

#### **Measurement of small neurons**

The neurons in the images of the immunostained trigeminal ganglion will be measured for the maximum diameter using Image J software. The neurons of diameter less than  $22\mu$  (Sankaran and Ramar, 2017) were small neurons, and the staining pattern of those neurons was studied.

### **RESULTS AND DISCUSSION**

[The G](#page-3-7)luR1 subunits was seen in the cytoplasm of neurons in the trigeminal ganglion (Figure 1). In migraine model rats, there was up-regulation of GluR1 subunits, and there was intense staining (Fig $ure 2$ ). The control rat staining was fine dots, and in migraine model rats, the staining was coar[se](#page-2-0) and granular. The most of the intense staining neurons the maximum diameter was less than  $22\mu$  proving mo[st](#page-2-1) of them are small neurons. The GluR1 was also localized in satellite glial cells and nerve fibers, but there was no change in the expression in the migraine model rats (Figure 1).

Peripheral stimulus leads to activation of AMPA receptors, especially GluR1 subunits, which causes a rapid influx of calcium inside the neurons. Followed by activation of protei[n k](#page-2-0)inases and transcription factors leading to changes in the membrane excitability of dendrites and alteration in cytoskeletal architecture of neurons. Until the calcium influx is prevented, this process is about to continue leading to excitotoxicity (Lu *et al.*, 2006), causing damage to neurons and creating pathogenesis for various disorders. Previous studies had shown the presence of glutamate receptors in primary afferent neurons, indicating their [involvement in](#page-3-8) the regulation of sensory processing. In this study, it was identified that the trigeminal ganglion neurons express GluR1, and these receptors increase its expression when there is nociceptive stimulation. Since AMPA receptors are ionotrophic glutamate receptors involved in fast glutaminergic transmission (Lu *et al.*, 2002) may be the reason for up-regulation as found in this study. As mentioned earlier, there are four subtypes of AMPA receptors, and all these receptors are studied in the sensory neurons. Th[e small neurons](#page-3-9) express GluR1, GluR2 and GluR3 and the GluR 2 and 3 expressions were mild compared to GluR1. The large neurons lack the expression of GluR1 subtype proving GluR1 is involved in nociceptive processing in the neurons which is supported by another study done in unmyelinated cutaneous axons for the

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**Figure 1: (a & b) : GluR1 localization large neurons (Red arrow), small neurons (Black arraow), satellite glial cell (Yellow arrow) and nerve ϐibers (White arrow) in control rats**

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**Figure 2: Migraine model rats showing intense localization of GluR1 in cytoplasm of the small neurons (Black arrow) and fine pale staining in large neurons (Blue arrow)** 

expression of AMPA receptors (Coggeshall and Carlton, 1998).

In the present study, the GluR1 was localized in all type of neurons (Large and Small), but the local[ization w](#page-3-10)as coarse in small neurons and fine in large neurons. Another study done on various types of glutamate receptors by recording the inward currents flowing through each of them following chronic constrictive injury showed the AMPA receptors are the major channels up-regulated during injury-causing largest inward current flowing proving that the AMPA receptors contribute most significantly to the nociception (Kopach *et al.*, 2011). This regulation can be explained based on calcium permeability through the GluR1during normal and injured state. Uninjured neurons, the GluR1 subunit of AMPA receptors is totally im[permeable to](#page-3-11) c[alcium](#page-3-11) ions with little permeability in GluR2 (Isaac *et al.*, 2007). Following any injury, there is a large proportion increase in GluR1 with an increase in permeability to calcium ions increasing plasti[city through](#page-3-12) activation of kinases resulting in early genes expression (Vikman *et al.*, 2008). The more specific activation of AMPA receptors resulted in mechanical allodynia and hyperalgesia and applying antagonist specific for calcium ion permeability reduced pain [\(Zhou](#page-3-13) *et al.*, 1[996\).](#page-3-13) The specific deletion of GluR1 subunit reduced pain and its sensitization in neuropathic pain models suggesting that GluR1 could exert pro-nociceptive action both in central and p[eripheral \(Gangadh](#page-3-14)aran *et al.*, 2011).

#### **CONCLUSION**

This study conc[ludes that the trigemina](#page-3-15)l ganglion neurons express GluR1 of AMPA receptors and these receptors expression increased following a migraine. This subunit up-regulation may be critical in pathogenesis of migraine since they are the fastest ionotrophic glutamate receptors. The GluR1 subunits were also expressed by satellite glial cells and nerve fibers, concluding they are also involved in the nociceptive sensitization. By developing antagonist to this subunit the migraine can be treated effectively so that the excitotoxicity can be prevented.

### **REFERENCES**

- <span id="page-3-0"></span>Bonsi, P., Cuomo, D., Persis, C. D., Centonze, D., Bernardi, G., Calabresi, P., Pisani, A. 2005. Modulatory action of metabotropic glutamate receptor (mGluR) 5 on mGluR1 function in striatal cholinergic interneurons. *Neuropharmacology*, 49:104– 113.
- <span id="page-3-10"></span>Coggeshall, R. E., Carlton, S. M. 1998. Ultrastructural analysis of NMDA, AMPA, and kainate receptors on unmyelinated and myelinated axons in the periphery. *The Journal of Comparative Neurology*, 391(1):78–86.
- Du, J., Zhou, S., Carlton, S. M. 2006. Kainate-induced excitation and sensitization of nociceptors in normal and inϐlamed rat glabrous skin. *Neuroscience*, 137(3):999–1013.
- <span id="page-3-15"></span>Gangadharan, V., Wang, R., Ulzhöfer, B., Luo, C., Bardoni, R., Bali, K. K., Todd, A. J. 2011. Peripheral calcium-permeable AMPA receptors regulate chronic inϐlammatory pain in mice. *The Journal of clinical investigation*, 121(4):1608–1623.
- <span id="page-3-12"></span>Isaac, J. T. R., Ashby, M. C., Mcbain, C. J. 2007. The Role of the GluR2 Subunit in AMPA Receptor Function and Synaptic Plasticity. *Neuron*, 54(6):859–871.
- <span id="page-3-1"></span>Julio-Pieper, M., Flor, P. J., Dinan, T. G., Cryan, J. F. 2011. Exciting Times beyond the Brain: Metabotropic Glutamate Receptors in Peripheral and Non-Neural Tissues. *Pharmacological Reviews*, 63(1):35–58.
- <span id="page-3-11"></span>Kopach, O., Kao, S. C., Petralia, R. S., Belan, P., Tao, Y. X., Voitenko, N. 2011. Inflammation alters trafficking of extrasynaptic AMPA receptors in tonically firing lamina II neurons of the rat spinal dorsal horn. *Pain*, 152(4):912–923.
- <span id="page-3-4"></span>Kung, L. H., Gong, K., Adedoyin, M., Ng, J., Bhargava, A., Ohara, P. T., Jasmin, L. 2013. Evidence for Glutamate as a Neuroglial Transmitter within Sensory Ganglia. *PLoS ONE*, 8(7).
- <span id="page-3-6"></span>Lee, C. J., Bardoni, R., Tong, C. K., Engelman, H. S., Joseph, D. J., Magherini, P. C., Macdermott, A. B. 2002. Functional Expression of AMPA Receptors on Central Terminals of Rat Dorsal Root Ganglion Neurons and Presynaptic Inhibition of Glutamate Release. *Neuron*, 35(1):729–737.
- <span id="page-3-5"></span>Lee, J., Ro, J. Y. 2007. Differential regulation of glutamate receptors in trigeminal ganglia following masseter inflammation. Neuroscience Letters, 421(2):91–95.
- <span id="page-3-8"></span>Lu, C., Wang, Y., Furukawa, K., Fu, W., Ouyang, X., Mattson, M. P. 2006. Evidence that caspase-1 is a negative regulator of AMPA receptor-mediated long-term potentiation at hippocampal synapses. *Journal of Neurochemistry*, 97(4):1104–1110.
- <span id="page-3-9"></span>Lu, C. R., Hwang, S. J., Phend, K. D., Rustioni, A., Valtschanoff, J. G. 2002. Primary afferent terminals in spinal cord express presynaptic AMPA receptors. *Journal of Neuroscience*, 22(21):9522–9529.
- <span id="page-3-3"></span>Pietrobon, D., Striessnig, J. 2003. Neurological diseases: neurobiology of migraine. *Nature Reviews Neuroscience*, 4(5):386–386.
- <span id="page-3-7"></span>Sankaran, P. K., Ramar, S. 2017. Morphological study of nociceptive neurons in the trigeminal ganglion. *Int J Health Rehab Sci*, 5(1).
- <span id="page-3-2"></span>Stone, J., Carson, A., Duncan, R., Roberts, R., Warlow, C., Hibberd, C., Sharpe, M. 2010. Who is referred to neurology clinics?-The diagnoses made in 3781 new patients. *Clinical Neurology and Neurosurgery*, 112(9):747–751.
- <span id="page-3-13"></span>Vikman, K. S., Rycroft, B. K., Christie, M. J. 2008. Switch to Ca 2+ -permeable AMPA and reduced NR2B NMDA receptor-mediated neurotransmission at dorsal horn nociceptive synapses during inflammatory pain in the rat. The Journal of Physi*ology*, 586(2):515–527.
- <span id="page-3-14"></span>Zhou, S., Bonasera, L., Carlton, S. M. 1996. Peripheral administration of NMDA, AMPA or KA results in pain behaviors in rats. *NeuroReport*, 7(4):895– 900.