



<https://ijrps.com>

ISSN: 0975-7538

Research Article

## ***In vitro* evaluation of phytochemical and antioxidant properties of *Syzygium cumini* leaves and their synergistic effect on its antimicrobial property**

Pranoti Belapurkar\* and Pragma Goyal

Department of Biotechnology, IPS Academy, Indore, Madhya Pradesh-452 012, India

### **ABSTRACT**

One of the major causes of ageing and stress-induced damage to cellular metabolism is release of Reactive Oxygen Species produced during normal metabolic processes. Plants have been reported to show antioxidant property attributed to their polyphenolic content. Thus, plant derived supplements should be incorporated in our daily diet as nutraceuticals. The methanolic extract of *Syzygium cumini* (L.) Skeels was evaluated for its phytochemical, antioxidant and antimicrobial properties. The extract showed the presence of tannins, alkaloids, flavonoids, saponins, terpenoids and glycosides. Total phenolic content, determined using gallic acid, was  $88.4 \pm 5.25$  mg/ml while total flavonoid content was found to be  $54.52 \pm 9.64$  mg/ml. Quercetin was used as standard. Its antioxidant property was determined by DPPH method, using ascorbic acid as standard. The methanolic extract was taken at concentrations ranging from 100-500 µg/ml. The percentage scavenging activity of the extract was found to be higher than the standard for the whole range. The antimicrobial property of the extract was determined by well-diffusion method. The extract showed potent antimicrobial activity against yeast and mold and Gram positive bacteria while the Gram negative bacteria were more resistant to the extract. The efficient antioxidant activity and antimicrobial potential suggests its use as dietary supplement to boost our immunity.

**Keywords:** DPPH; nutraceutical; reactive oxygen species; total flavonoid content; total phenolic content; well-diffusion method

### **INTRODUCTION**

There are different free radical scavenging molecules like superoxide dismutase, catalase, peroxidase and glutathione present naturally in our body. Reactive Oxygen Species (ROS) viz., singlet oxygen ( $O^1$ ), superoxide ion ( $O_2^-$ ), hydroxide ion ( $OH^-$ ) and hydrogen peroxide ( $H_2O_2$ ) are produced during normal metabolic processes and cause oxidative damage to biomolecules like DNA and protein (Halliwell, 1997). These ROS are reportedly responsible for many grave human diseases eg cancer (Parmar et al., 2010), Alzheimer's, diabetes, Parkinson's etc (Ozgen et al., 2006). To counter the effects of ageing and stress-induced damage to cellular metabolism, it is imperative to add plant derived dietary supplements that boost our mechanism of scavenging ROS (Battacharya et al., 1997; Ilavarasan et al., 2001; Manonmani et al., 2002; Li et al., 2008).

Ancient Indian medicinal literature like *Sushrut Samhita* and *Charak Samhita* have emphasized on the therapeutic use of plant extracts. The active components of

different plants possess high antioxidant properties which make them effective scavengers of ROS and potent antimicrobial agents. Different plant parts contain polyphenolic compounds like flavonoids, tannins and tocopherol which are responsible for their antioxidant properties (McCord, 2000). *Syzygium cumini* (L.) Skeels, a plant of Myrtaceae family, is a tropical evergreen tree, with its origins in India. The earlier reported work on this plant has been majorly on its fruits and seeds (Banerjee and Narendhirakannan, 2011; Borhade, 2012; Murti et al., 2012; Saha et al., 2013). These parts have been reported for their antioxidant, anti-inflammatory (Muruganandan et al., 2001), neuro-psychopharmacological, antimicrobial (Bhuiyan et al., 1996; Shafi et al., 2002), anti-HIV, antileishmanial (Chandrasekaran et al., 2004; Ratnam et al., 2008), nitric oxide scavenging (Jagetia et al., 2002), free radical scavenging (Silva et al., 2006), antidiarrhoeal (Mukherjee et al., 1998), antifertility (Rajasekaran et al., 1988), gastro-protective, antiulcerogenic and radio-protective activities (Sagrawat et al., 2006). There is a wide scope to study the medicinal properties of its leaves like antimicrobial and antioxidant activity and correlate it with its secondary metabolites, which is the focus of this study.

### **MATERIALS AND METHODS**

#### **Plant source**

Fresh mature leaves of *S. cumini* were collected locally from Indore City, Madhya Pradesh in the month of July.

\* Corresponding Author

Email: pranotivivek@gmail.com

Contact: +91-9425307608

Received on: 30-09-2014

Revised on: 05-11-2014

Accepted on: 11-11-2014

They were cleaned, air dried, deveined and pulverized into fine powder and stored in an air tight container at room temperature.

### Preparation of extract

For extract preparation, 10gms of leaf powder was defatted using 250 ml petroleum ether (Boiling Point 40-60°C) followed by methanolic extraction using Soxhlet apparatus. The extract was then vacuum dried and stored at 4°C in air tight containers for further use.

#### 1. Qualitative Phytochemical Analysis

Tannins, phenols, alkaloids, flavonoids, glycosides, saponins and steroids were analyzed qualitatively by using standard protocols (Harborne et al., 1973; Trease et al., 1989).

#### 2. Quantitative Phytochemical Analysis

Total phenolic content was estimated spectrophotometrically using Gallic acid as standard, by Folin-Ciocalteu method modified by Singleton et al., 1999. Total flavonoid content was determined using the methodology of Jia et al., 1999 with slight modifications. Quercetin was used as standard. All the determinations were carried out in triplicates.

#### 3. Antioxidant assay

To assay the antioxidant property, DPPH free radical scavenging activity (Chang et al., 2002) was performed. Ascorbic acid was used as standard. The stock solution of extract (1mg/ml) was diluted to final concentrations ranging from 100-500 µg/ml in methanol. To 2.5ml of each sample concentration, 1 ml of 0.3mM DPPH solution was added. Blank was made using methanol and negative control had DPPH (1ml) and methanol (2.5ml). After reaction for 30mins, the absorbance was measured at 518nm. Similar procedure was followed for the standard. All the procedures were carried out in triplicates. The percentage scavenging activity was calculated by,

$$\% \text{ Scavenging activity} = 100 - \left[ \frac{\text{Absorbance of sample}}{\text{Absorbance of blank}} \times \frac{\text{Absorbance of control}}{\text{Absorbance of blank}} \right]$$

#### 4. Antimicrobial activity

*In vitro* antimicrobial activity of methanolic extract of *S. cumini* leaves was determined against few Gram positive and Gram negative bacteria and fungal cultures. The standard cultures were procured from NCIM, Pune (Table No.2). They were sub-cultured on MRS medium and stored under refrigerated conditions till further use.

For testing, 24 hrs old cultures were inoculated on Nutrient Agar plates for bacterial cultures and on SDA plates for fungal cultures. The antimicrobial activity was determined by well diffusion method (Perez et al., 1990). The test cultures were lawn cultured and 100-200µl of extract was added to wells aseptically. The

plates were incubated for 24 hrs at 37°C for bacterial cultures and for 48-72 hrs at 28°C for fungal cultures.

## RESULTS AND DISCUSSION

### 1. Qualitative analysis

The phytochemical analysis showed the presence of alkaloids, flavonoids, tannins, phenols, steroids and saponins (Table No.1). These compounds are known to have beneficial importance in medicine. Flavonoids have been reported by different workers as an anti-allergic, antiinflammatory, antimicrobial and anticancerous agent (Aiyelaagbe and Osamudiamen, 2009). Yoshizawa et al., 1987 and Okuda et al., 1992 have earlier reported activities like antiviral, antibacterial and anti-tumor for tannins. Saponins are surface active agents and therefore allow antibody access in intracellular proteins. In medicine they have reportedly being used in hyperglycemia, hypercholesterolemia, weight loss etc (Gowri and Vasantha, 2010).

### 2. Quantitative analysis

The total phenolic content of methanolic extract of *S. cumini* leaves was 88.4±5.25 mg/gm. The results indicate that the total phenolic content was significantly high suggesting that methanolic extract of *S. cumini* leaves possess higher amounts of secondary metabolites. This further confirms that phenolic compounds are better extracted in polar solvents like methanol and ethanol as compared to aqueous extraction (Sultana et al., 2009).

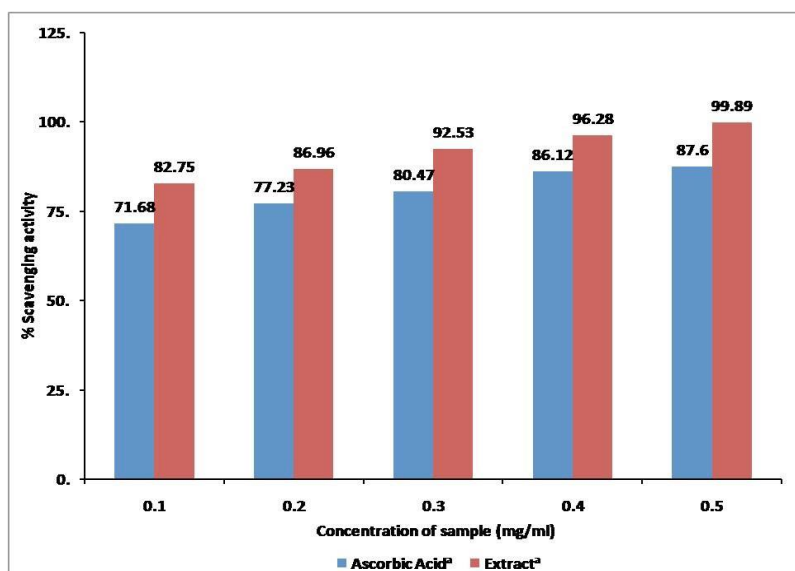
Reports obtained earlier have focused greatly on the free radical scavenging activity of the phenolics. They are produced as secondary metabolites in plants and chelate transitional metals and scavenge free radicals; thus they work as good antioxidants (Mohamed et al., 2010).

*S. cumini* methanolic leaf extract showed significantly high total flavonoid content of 54.52± 9.64 mg/gm. Earlier reports have emphasized the antioxidant properties of flavonoids (Zheng et al., 2011). These compounds possess certain hydroxyl functional groups which impart antioxidant effect through either chelating or scavenging mechanisms (Das and Pereria, 1990; Kessler et al., 2003). The present study confirms high flavonoid content in methanolic extract of *S. cumini* leaves, thus suggesting its high antioxidant effect. This further suggests its use in human nutrition for considerable health benefits.

The present work further corroborates with previous reports that the antioxidant potential of a plant is directly proportional to its total phenolic and flavonoid content (Mohamed et al., 2013).

### 3. Antioxidant activity

The different increasing concentrations of the methanolic extract of *S. cumini* leaves showed increment in the free radical scavenging activity with the increase in



\*The value is mean of three determinants

Figure 1: DPPH scavenging activity of standard and methanolic extract of *S. cumini* leaves

Table 1: Phytochemical constituents of methanolic extract of *S. cumini* leaves

S.No.	Phytochemical constituents	Methanolic Extract of <i>S.cumini</i>
1.	Alkaloids	+
2.	Flavonoids	+
3.	Tannins	+
4.	Saponins	+
5.	Terpenoids	+
6.	Glycosides	+

Table 2: Antimicrobial activity of methanolic extract of *S. cumini* leaves

S.no.	Name of microorganisms	NCIM No.	Diameter of zone of inhibition (mm)
1.	<i>Staphylococcus aureus</i>	2079	19
2.	<i>Escherichia coli</i>	2065	17
3.	<i>Bacillus subtilis</i>	2063	16
4.	<i>Streptococcus faecalis</i>	5024	---
5.	<i>Candida albicans</i>	3471	23
6.	<i>Aspergillus niger</i>	1196	20

the extract concentration (Figure 1). There was significant decrease in the colour of DPPH free radical indicating scavenging activity of methanolic extract. The scavenging capability of the extract and ascorbic acid, used as standard, was found to be significant ( $P \leq 0.05$ ). These results are in sync with the high total phenolic content in the extract. The phenolics with their redox properties have the ability to chelate transitional metals, scavenge free radicals, inhibit lipooxygenase and show good antioxidant properties (Decker, 1997; Naik et al., 2003; Mohamed et al., 2013).

Apart from phenolics, the extract is rich in polar phytochemicals like flavonoids, tannins etc., which have been extracted efficiently by polar solvent, methanol in this study. Therefore the extract has shown good antioxidant activity which corroborates with the previous works of Sharma et al., 2003; Reynertson et al., 2008; Al-Reza et al., 2009; Mohamed et al., 2013.

#### 4. Antimicrobial activity

The methanolic extract of *S. cumini* showed good antifungal and antibacterial activity *in vitro*. The results showed highest zone of inhibition against yeast and mould; *C. albicans* was comparatively more sensitive than *A. niger*. Good sensitivity was observed for *S. aureus* followed by *E. coli* and *B. subtilis*. *S. faecalis* was found to be resistant to the extract (Table No.2). This suggests that the extract is significantly effective against Gram positive bacteria than Gram negative bacteria. This antimicrobial property can be attributed to the synergistic effect of high total phenolic and total flavonoid content of the extract, which are reportedly good antioxidants (Gowri and Vasantha 2010; Mohamed et al., 2013).

#### CONCLUSION

The current study has shown that the total phenolic content and total flavonoid content of the methanolic

extract of *S. cumini* leaves was very high suggesting it to be a good antioxidant. This high polyphenolic content is also responsible for its potent antifungal and antibacterial activity. Therefore it is recommended that this extract should be incorporated in daily diet as a nutraceutical supplement.

#### CONFLICT OF INTEREST

No conflict of interest lies between Authors

#### ACKNOWLEDGEMENTS

The authors would like to acknowledge Dr. Sanjay Nagar, Head of Department, Biotechnology and Ar. Achal Choudhary, President IPS Academy, Indore for guidance and financial assistance provided for the study.

#### REFERENCES

- Aiyelaagbe, O.O. and Osamudiamen, P.M. Phytochemical screening for active compounds in *Mangifera indica*, *Plant Sciences Research*, vol. 2, no. 1, 2009 pp. 11-13.
- Al-Reza, S.M., Rahman, A. and Kang, S.C. Chemical composition and inhibitory effect of essential oil and organic extracts of *Cestrum nocturnum* L. on food-borne pathogens, *International Journal of Food Science and Technology*, vol. 44, 2009 pp. 1176-1182.
- Banerjee, J. and Narendhirakannan, R.T. Phytochemical analyses, antibacterial, in vitro antioxidant and cytotoxic activities of ethanolic extract of *Syzygium cumini* (L.) seed extract, *International Journal of Pharmaceutical Sciences and Research*, vol. 2, no.7, 2011 pp. 1799-1806.
- Battacharya, S. K., Satyan, K. S. and Ghosal, S. Antioxidant activity of glycowithanolides from *Withania somnifera*, *Indian Journal of Experimental Biology*, vol. 35, 1997 pp. 236-239.
- Bhuiyan, M.S.A., Mia, M.Y. and Rashid M.A. Antibacterial principles of the seeds of *Eugenia jambolana*, *Bangladesh Journal of Botany*, vol. 25, no. 2, 1996 pp. 239-241.
- Borhade, S. Antibacterial activity, phytochemical analysis of water extract of *Syzygium cumini* and analytical study by HPLC, *Asian Journal of Experimental Biological Sciences*, vol. 3, no. 2, 2012 pp. 320-324.
- Chandrasekaran, M. and Venkatesalu, V. Antibacterial and antifungal activity of *Syzygium jambolanum* seeds, *Journal of Ethnopharmacology*, vol. 91, 2004 pp. 105-108.
- Chang, W., Choi, C.K.S., Hwang, S.S., Bong, K.C., Hye, J.A., Min, Y.L., Sang, H.P. and Kim, S.K. Antioxidant activity and free radical scavenging capacity between Korean medicinal plants and flavonoids by assay guided comparison, *Plant Science*, vol. 163, 2002 pp. 1161-1168.
- Das, N.P. and Pereira, T.A. Effects of flavonoids on thermal autooxidation of palm oil: structure activity relationship, *Journal of American Oil Chemists' Society*, vol. 67, 1990 pp. 255-258.
- Decker, E.A. Phenolics: prooxidants or antioxidants?, *Nutrition Reviews*, vol. 55, 1997 pp. 396-407.
- Gowri, S.S. and Vasantha, K. Phytochemical Screening and Antibacterial Activity of *Syzygium cumini* (L.) (Myrtaceae) Leaves Extracts, *International Journal of PharmTech Research*, vol. 2, no. 2, 2010 pp. 1569-1573.
- Halliwell, B. Antioxidants and human disease: a general introduction, *Nutrition Reviews*, vol. 55, no.1, 1997 pp. 44-52.
- Harborne, J.B. *Phytochemical methods, a guide to modern techniques of plant analysis*, Chapman and Hall, London, 1973.
- Ilavarasan, R., Mohideen, S., Vijayalakshmi, M. and Manonmani, G. Hepatoprotective effect of *Cassia angustifolia* Vahl, *Indian Journal of Pharmaceutical Sciences*, vol. 63, 2001 pp. 504-507.
- Jagetia, G.C. and Baliga, M.S. *Syzygium cumini* (Jamun) reduces the radiation-induced DNA damage in the cultured human peripheral blood lymphocytes: a preliminary study, *Toxicology Letters*, vol. 132, 2002 pp. 19-25.
- Jia, Z., Tang, M. and Wu, J. The determination of flavonoid content in mulberry and their scavenging effects on superoxide radicals *Food Chemistry*, vol. 64, 1999 pp. 555-599.
- Kessler, M., Ubeaud, G., and Jung, L. Anti- and prooxidant activity of rutin and quercetin derivatives. *Journal of Pharmacy and Pharmacology*, vol. 55, 2003 pp. 131-142.
- Li, H.B., Wong, C.C., Cheng, K.W. and Chen, F. Antioxidant properties in vitro and total phenolic contents in methanol extracts from medicinal plants, *LWT - Food Science and Technology*, vol. 41, 2008 pp. 385-390.
- Manonmani, G., Anbarasi, K., Balakrishna, K., Veluchamy, G. and Shyamala Devi, C. S. Effect of *Terminalia arjuna* on the antioxidant defence system in alloxan induced diabetes in rats, *Biomedicine*, vol. 22, 2002 pp. 52-61
- McCord, J.M. The evolution of free radicals and oxidative stress, *The American Journal of Medicine*, vol. 108, 2000 pp. 652-659.
- Mohamed, A.A., Khalil, A.A. and El-Beltagi, H.E.S. Antioxidant and antimicrobial properties of kaff maryam (*Anastatica hierochuntica*) and doum palm (*Hyphaene thebaica*), *Grasas Y Aceites*, vol. 61, no. 1, 2010 pp. 67-75.

- Mohamed, A.A., Ali, S.I. and El-Baz, F.K. Antioxidant and antibacterial activities of crude extracts and essential oils of *Syzygium cumini* leaves, *Plos One*, vol. 8, no. 4, 2013 pp. 1-7.
- Mukherjee, P.K., Saha, K., Murugesan, T., Mandal, S.C., Pal, M. and Saha, B.P. Screening of anti-diarrhoeal profile of some plant extracts of a specific region of West Bengal India, *Journal of Ethnopharmacology*, vol. 60, 1998 pp. 85-89.
- Murti, K., Paliwal, D., Madan, S., Kundu, R. and Kaushik, M. Exploration of preliminary phytochemical studies of seeds of *Syzygium cumini*, *American Journal of Pharmacology and toxicology*, vol. 7, no. 1, 2012 pp. 12-14.
- Muruganandan, S., Srinivasan, K., Chandra, S., Tandan, S.K., Lal, J. and Raviprakash, V. Anti-inflammatory activity of *Syzygium cumini* bark, *Fitoterapia*, vol. 72, 2001 pp. 369-375.
- Naik, G.H., Priyadarsini, K.I., Satav, J.G., Banavalikar, M.M., Sohani, D.P., Biyani, M.K. and Mohan, H. Comparative antioxidant activity of individual herbal components used in ayurvedic medicine, *Phytochemistry*, vol. 63, 2003 pp. 97-104.
- Okuda, T., Yoshida, T. and Hatano, T. Polyphenols from Asian plants, structural diversity and antitumor and antiviral activities, in Huang, M.-T., Ho, C.-T., Lee, C.Y. (ed.) *Phenolic Compounds in Food and Their Effects on Health II, Antioxidants and Cancer Prevention*, Washington DC, USA: American Chemical Society, 1992.
- Ozgen, U., Mavi, A., Terzi, Z., Yildirim, A., Coskun, M and Houghton, P. J. Antioxidant properties of some medicinal Lamiaceae species, *Pharmaceutical Biology*, vol. 44, 2006 pp. 107-112.
- Parmar, J., Sharma, P., Verma, P., Sharma, P. and Goyal, P.K. Chemopreventive action of *Syzygium cumini* on DMBA-induced skin papillomagenesis in mice, *Asian Pacific Journal of Cancer Prevention*, vol. 11, 2010 pp. 261-265.
- Perez, C., Paul, M. and Bazerque, P. An Antibiotic assay by the agar well diffusion method, *Acta Biologicae et Medecine Experimentalis*, vol. 15, 1990 pp. 113-115.
- Rajasekaran, M., Bapna, J.S., Lakshmanan, S., Nair, R.A.G., Veliath, A.J. and Panchanadam, M. Antifertility effect in male rats of oleanolic acid a triterpene from *Eugenia jambolana* flowers, *Journal of Ethnopharmacology*, vol. 24, 1988 pp. 115-121.
- Ratnam, K.V. and Raju, R.R.V. In vitro antimicrobial screening of fruit extract of two *Syzygium* species (Myrtaceae), *Advances in Biological Research*, vol. 2, no. 1-2, 2008 pp. 17-20.
- Reynertson, K.A., Yang, H., Jiang, B., Basile, M.J. and Kennelly, M.E.J. Quantitative analysis of antiradical phenolic constituents from fourteen edible Myrtaceae fruits. *Food Chemistry*, vol. 109, no. 4, 2008 pp. 883-890.
- Sagrawat, H., Mann, A.S. and Kharya, M.D. Pharmacological potential of *Eugenia jambolana*: a review, *Pharmacognosy Magazine*, vol. 2, 2006 pp. 96-104.
- Saha, R.K., Zaman, N.M. and Roy, P. Comparative evaluation of medicinal activities of methanolic extract of seeds, fruit pulps and fruit juice of *Syzygium cumini* in vitro, *Journal of Coastal Life Medicine*, vol. 1, no. 4, 2013 pp. 300-308.
- Shafi, P.M., Rosamma, M.K., Jamil, K. and Reddy, P.S. Antibacterial activity of *Syzygium cumini* and *Syzygium travancoricum* leaf essential oils, *Fitoterapia*, vol. 73, no. 5, 2002 pp. 414-416.
- Sharma, S.B., Nasir, A., Prabhu, K.M., Murthy, P.S. and Dev, G. Hypoglycaemic and hypolipidemic effect of ethanolic extract of seeds of *Eugenia jambolana* in alloxan-induced diabetic rabbits, *Journal of Ethnopharmacology*, vol. 85, no. 2-3, 2003 pp. 201-206.
- Silva, D.H.S., Plaza, C.V., Bolzani, V.S., Cavalheiro, A.J. and Castro-Gamboa, I. Antioxidants from fruits and leaves of *Eugenia jambolana*, an edible Myrtaceae species from Atlantic Forest, *Planta Medica* vol. 72, 2006 pp. 1038.
- Singleton, V.L., Orthofer, R. and Lamuela-Raventos, R.M. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent, *Methods in Enzymology*, vol. 299, 1999 pp. 152-178.
- Sultana, B., Farooq, A. and Muhammad, A. Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts, *Molecules*, vol. 14, 2009 pp. 2167-2180.
- Trease, G.E. and Evans, W.C. A text book of pharmacognosy, London: Academic press, 1989.
- Yoshizawa, S., Horiuchi, T., Fujiki, H., Yoshida, T., Okuda, T. and Sugimura, T. Antitumor promoting activity of (-)-Epigallocatechin gallate, the main constituent of "Tannin" in green tea, *Phytotherapy Research*, vol. 1, 1987 pp. 44-47.
- Zheng, N., Wang, Z., Chen, F. and Lin, J. Evaluation to the antioxidant activity of total flavonoids extract from *Syzygium jambos* seeds and optimization by response surface methodology, *African Journal of Pharmacy and Pharmacology*, vol. 5, no. 21, 2011 pp. 2411-2419.