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# Ventilation tube a fleeting alcove for multi drug resistant non-hemolytic Staphylococcus epidermidis

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### ABSTRACT

Health care associated infections are escalating globally. Ventilation tube is one of the potential reservoirs for microorganisms. In view of this a prospective study was undertaken to detect the microbial spectrum, haemolytic assay and antibiotic susceptibility of nosocomial bacteria on ventilation tube. Samples were collected by wiping the sterile moist swab over the ventilation tube in intensive care unit (ICU). Collected samples were subjected to standard isolation and characterization procedures. *Staphylococcus epidermidis* was isolated from the collected samples. For hemolytic assay, loopful of *Staphylococcus epidermidis* culture was streaked on freshly prepared blood agar and incubated overnight. Hemolytic assay revealed non-hemolytic *Staphylococcus epidermidis*. Antimicrobial susceptibility test was performed according to CLSI (2007). Bacterial isolates were subjected to disc diffusion test against gentamicin (120µg), vancomycin (30µg), erythromycin (15µg) tetracycline (30µg), ampicillin (10µg), amoxicillin (10µg) doxycycline (30µg), bacitracin (10µg) and ceftazidime (30µg). Antimicrobial susceptibility test revealed Staphylococcus *epidermidis* sensitivity to gentamycin and vancomycin but resistant to all antibiotics tested.

Keywords: Health care associated infection; Commensals; Skin infection; Antibiotic susceptibility.

### INTRODUCTION

Health care associated infections are recognized as public health predicament worldwide and the infections are even more startling in the 21<sup>st</sup> century (Samuel S.O. 2010). The world health organization (WHO) estimates 190 million hospitalized patients annually worldwide, 9 million patients contact health care associated infections and 1 million patients succumb to death due to health care associated infections (Paris: C-CLIN Paris-Nord, 1995). Staphylococcus epidermidis is a normal inhabitant of human skin frequently known to cause foreign body associated infections (Rupp.M.E et al., 1994; Kloos WE et al., 1975). The pathogenesis of S.epidermidis infection is linked with the ability to form biofilm on polymer surface such as ventilation tube (Lucilla B et al., 1997). Mechanical ventilators are associated with hospital acquired infections and the disease occurs 10 to 20% more to patients who are on ventilator longer than 48 hours and is associated with significant increase in length of stay in hospital, mortality and cost (Jarvis WR 2007). Since the infections associated with S. epidermidis are chiefly acquired during hospitalization, it is not surprising that resistant pathogens

\* Corresponding Author Email: pallavijayavanth@gmail.com Contact: Received on: 21-07-2011 Revised on: 06-09-2011 Accepted on: 07-09-2011 pose an amplified challenge to the hospital for prevention and treatment (Bradley JS et al., 2007).

### METHODOLOGY

Surface samples were collected by wiping the surface of selected areas with sterile cotton swab. Sampling procedures was done according to CDC (2003). All collected samples were transported to Research Laboratory for isolation, characterization and antibiotic susceptibility test according to standard procedures (Baron J & Finglod S., 2007; CLSI 2007).

A standard inoculum adjusted to 0.5 McFarland was swabbed on Muller – Hinton agar and antibiotic disc of gentamicin (120µg), vancomycin (30µg), erythromycin (15µg) tetracycline (30µg), ampicillin (10µg), amoxicillin (10µg) doxycycline (30µg), bacitracin (10µg) and ceftazidime (30µg) were placed at equidistance after drying the plate for 2-4 min and incubated at 37°C for 24 hours and the zone of inhibition was measured and compared with standard antibiotic susceptibility chart (CLSI, 2007).

### RESULTS

*Staphylococcus epidermidis* was the predominant bacteria isolated from the ventilation tube in Intensive care Unit (ICU). It was present on all the ventilation tubes that were used in this study.

The results of the antibiotic susceptibility test are shown in table 1. *S.epidermidis* was found to be sensitive to gentamycin and vancomycin. On the other

Groups	Antibiotic agent	Potency (µg)	Inhibition zone (mm)	S/I/R
Aminoglycoside	CN	120	25	S
Glycopeptides	VA	30	22	S
Penicillin	E	15	9	R
	AMP	10	14	R
	AML	10	14	R
Tetracycline	DO	30	11	R
	TE	30	10	R
Cephalosporin	CAZ	30	7	R
Others	В	10	14	R

Table 1: Antibiotic susceptibility of Staphylococcus epidermidis against antibiotics

S: sensitive, I: Intermediate, R: resistant. CN: Gentamycin, VA: Vancomycin, E: Erythromycin, TE: Tetracycline, AMP: Ampicillin, AML:Amoxicilin, DO: Doxycycline, B: Bacitracin, CAZ: Ceftazidime.

hand, *S.epidermidis* was resistant to erythromycin, tetracycline, ampicilin, amoxicillin, doxycycline, bacitracin and ceftazidime.

Ceftazidime showed the least inhibition zone (7mm), followed by erythromycin (9mm), tetracycline (10mm), doxycycline (11mm), ampicilin, amoxicillin and bacitracin (14mm) respectively. Where as inhibition zone for gentamycin was 25mm and 22mm forvancomycin which is the indicator of sensitivity.

#### DISCUSSION

*S.epidermidis* is commonly found on human skin that sometimes causes human illness. They are usually dismissed as contaminant but now it is the most important bacteria causing nosocomial infection especially in individuals with weakened immune system, newborn children and people with implanted medical devices (D Isaacs. 2003). Difficulty in distinguishing the *S.epidermidis* strain is commonly encountered by researchers (Von Eiff C et al., 2001). According to our findings *S.epidermidis* residing on the ventilation tube is likely to infect other anatomical sites of the same patient during medical procedures via various modes of transmission. However, this does not hold true in every case due to many influencing factors.

The significant finding of this study is isolation of gram positive *S.epidermidis* from ventilation tube contrast to gram negative bacteria causing pneumonia reported by researcher (P G Flanagan et al., 2001). This indicates the moist environment of ventilation tube bolsters the proliferation of *S.epidermidis* (M Dwivedi et al., 2009). *S.epidermidis* was isolated from the outer surface of ventilation tube hence *S.epidermidis* could be considered transient bacteria however, periodic screening will aid in determining if ventilation tube continuously harbours *S.epidermidis* which could have been originated from various sources namely medical personnel, patient attendants and contaminated air, in addition inadequate decontamination protocol could also be the governing factor.

Antibiotic susceptibility test revealed a contrast finding compared to other findings (Denise O. Garrett et al, 1999). This study showed *S.epidermidis* sensitive to

vancomycin and gentamycin and resistant to ampicilin, amoxicillin, erythromycin, tetracycline and doxy- cycline.

The best method to reduce the frequency of resistant bacteria is to use antibiotic rotation or cycling (Jean-Louis Vincent. 2003). The choice of antibiotic treatment can affect the bacterial spectrum of the entire hospital community. Infections caused by *S.epidermidis* are often persistent and relapsing especially it is correlated with the ability to form biofilm on polymer surface (Valentin-weigand P et al., 1993). In addition, biofilm infections often give recurring symptoms until proper therapy is given. The most inexpensive method available is to break the chain of infection to implement the same identifying the epidemiological factors are imperative CDC (2002) Further, strict implementation of sterilization and disinfection protocols (Rutala MW 2004)

### CONCLUSION

Continuous education on infection control should be advocated. Hospital-wide campaigns and screening should be done periodically and remind hospital personnel on better implementation of existing infection control guidelines to reduce the probability of health care associated infections caused especially by antibiotic resistant bacteria.

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### REFERENCES

- Baron J, Finglod S. Methods for identification of etiologic agents of infectious diseases. In: Betty A, Forbers F (eds), Baily & Scott's diagnostic microbiology. 12th ed, St. Louis, USA, Mosby, Inc, 2007; 300-29.
- CDC guideline for hand hygiene in health- care setting: Recommendation of the health care infection control practices advisory committee and the HICPAC/SHEA

hand hygiene task force. MMWR. Vol. 51, 2002 pp. 1-4.

- CDC guidelines for environmental infection control in health-care facilities: recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee (HICPAC). MMWR. Vol. 52, 2003 pp 1-48.
- Centre de coordination de la lutte contre les infections nosocomiales de l'inter région Paris et Nord. Guide de definition des infections nosocomiales. Paris: C-CLIN Paris-Nord, 1995. From <a href="http://www.caducee.net/DossierSpecialises/infection/nosocomiales.asp">http://www.caducee.net/DossierSpecialises/infection/nosocomiales.a sp > (Retrieved April 4, 2011).</a>
- D Isaacs, on behalf of the Australasian Study Group for Neonatal Infections. A ten year, multicentre study of coagulase negative staphylococcal infections in Australasian neonatal units. Arch Dis Child Fetal Neonatal Ed. Vol. 88, 2003 pp. 89–93.
- Denise O. Garrett, Elise Jochimsen, Kate Murfitt, Bertha Hill, Sigrid McAllister, Pat Nelson, Richard V. Spera, Richard K. Sall, Fred C. Tenover, Judy Johnston, Barbara Zimmer, and William R. Jarvis. The Emergence of Decreased Susceptibility to Vancomycin in *Staphylococcus epidermis.* Infection Control and Hospital Epidemiology. Vol. 20, no.3, 1999 pp 167-170.
- Jarvis, WR. The Lowbury Lecture. The United States approach to strategies in the battle against healthcare-associated infections, 2006: transitioning from benchmarking to zero tolerance and clinician accountability. Journal Hospital Infection. Vol. 65, no.2, 2007 pp. 3-9.
- Jean-Louis Vincent. Nosocomial infection in adult intensive-care units. The Lancet. Vol. 361, 2003 pp. 2068-2077.
- John S Bradley, Robert Guidos, Steve Baragona, John G Bartlett, Ethan Rubinstein, George G Zhanel, Michael D Tino, David L Pompliano, Frank Tally, Praveen Tipirneni, Glenn S Tillotson, John H Powers, Glenn S Tillotson. Anti-infective research and development problems, challenges, and solutions. The Lancet Infectious Diseases. Vol. 7, no.1, 2007 pp. 68-78.
- Kloos WE, Musselwhite MS. Distribution and persistence of Staphylococcus and Micrococcus species and other aerobic bacteria on human skin. Appl Microbiol. Vol. 30, no.3, 1975 pp. 381-385
- Lucilla Baldassarri, Gianfranco Donelli, Antonella Gelosia, Andrew W. Simpson and Gordon D. Christensen. Expression of Slime Interferes with In vitro detection of Host Protein Receptors of Staphylococcus epidermidis. Infection and Immunity. Vol. 65, no.4 1997 pp. 1522-1526.
- M Dwivedi, A Mishra, RK Singh, A Azim, AK Baronia, KN Prasad. Nosocomial cross-transmission of Pseudomonas aeruginosa between patients in a tertiary in-

tensive care unit. Indian J Pathol Microbiol. Vol. 52, 2009 pp. 509-13.

- P G Flanagan, S K Jackson, G Findlay. Diagnosing of Gram negative, ventilator associated pneumonia by assaying endotoxin in bronchial lavage fluid. Journal Clinical Pathology. Vol. 54, 2001 pp. 107-110.
- Rupp ME, Archer GL. Coagulase negative staphylococci: pathogen associated with medical progress. Clinical Infectious Disease. Vol.19, no.2, 1994 pp. 231-243.
- Rutala MW, Weber DJ. Disinfection and sterilization in health care facilities: what clinicians need to know? Clin Infect Dis, Vol.39, 2004 pp. 702-9
- Samuel, S.O, Kayode.O.O, Musa, O.I, Nwigwe,G.C, Aboderin A.O, Salami T. A.T, taiwo S.S. Nosocomial infection and the challenges of control in developing countries. African journal of clinical and experimental microbiology. Vol.11, no.2, 2010 pp. 102-110.
- Valentin-Weigand P, Timmis KN, Chhatwal GS. Role of fibronectin in staphylococcal colonisation of fibrin thrombi and plastic surfaces. Journal Medical Microbiology. Vol.38, no.2, 1993 pp. 90-95.
- Von Eiff C, Becker K, Machka K. Nasal carriage as a source of Staphylococcus aureus bacteremia. New England Journal of Medicine. Vol. 344, 2001 pp. 11–16.