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Molecular docking studies of pancreatic cancer expressed proteins with *Psidium guajava* **(guava) derived bioactive compounds**

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INTRO[DUCTION](https://ijrps.com)

Cancer is not just the accumulation of tissue masses but also the heterotypic interaction of distinct cell types with one another (Hanahan and Weinberg, 2011). Cancer was the second leading cause of mortality in 2018, resulting in 9.6 million deaths

(WHO, https://www.who.int/health-topics/cancer #tab=tab_1). Pancreatic cancer is the fourth leading cause of cancer deaths in the united states and this is because the annual death rate being equal to the annual incidence rates (Street, 2019). Pancreatic cancer has a survival rate of only 5 years that too the percent of survival rate in that is only 9 % (Rahib *et al.*, 2014). The reason behind this is the inability of the disease to be [even diagnos](#page-7-0)ed and cured (Warshaw and del Castillo, 1992). It is found that 80 % of pancreatic cancers are unresectable in patie[nts and also](#page-7-1), [the s](#page-7-1)urvival rate of the subjects after surgery is poor (Rahib *et al.*, 2014). Pancreatic cancer [is estimated to be the second lead](#page-7-2)ing cause of death by 2030 (Gilabert *et al.*, 2017). Among all the other cancer types, pancreatic cancer exhibited high resistance to traditio[nal chemotherapy](#page-7-1) and radiation therapy, including both de novo (intrinsic) and acquired (thera[py-induced\) chemo-r](#page-6-1)esistance of the cancer cells (Long *et al.*, 2011). The development of drug resistance is a critical barrier for the effective transportation of a drug to the target site in the tumor. There are two types of drug resistance, the one being de novo resistance and the other acquired resistance. De novo drug resistance subjects do not respond to the chemotherapy from the beginning of its administration and acquired resistance subjects are first sensitive to the drug but later, they become resistant to the treatment. The cancer cells also become resistant to other mechanically and structurally unrelated drugs, which is called multidrug resistance (Jäger, 2009). Thus, a treatment combining several drugs for different targets also fail (Wang *et al.*, 2011; Gottesman *et al.*, 2002).

Currently, there are many combinational therapies for pancre[atic cancer.](#page-6-2) Gemcitabine is used as the drug of choice for enfeebled patients and in [com](#page-7-3)[bination al](#page-7-3)[ong with other drugs](#page-6-3) such as irinotecan, oxaliplatin (also known as FOLFIRINOX), 5 fluorouracil, leucovorin in the case of healthy subjects (Kobayashi *et al.*, 2017; Orlandi *et al.*, 2016). But due to the short half-life range of 8 to 15 mins for the drug, the continuous administration of the drug parenterally causes renal and hematological toxicities([Dorjee and Long](#page-6-4), [2018](#page-6-4); [Kasuya](#page-7-4) *et al.*, [2011](#page-7-4); Muranaka *et al.*, 2017). Although gemcitabine is used as a first-line drug, it is not satisfactory due to the endogenous and exogenous resistance exhibited. [Endogenous drug](#page-6-5) r[esista](#page-6-5)[nce occurs due to the](#page-6-6) [drug metabolism](#page-6-7), [its tra](#page-6-7)nsport, abnormal activation and inactivation of its signaling pathways, whereas exogenous drug resistance occurs due to impedance of drug delivery to the target (Hung *et al.*, 2012; Nakano *et al.*, 2007).

Psidium Guajava (commonly called as guava) is a native fruit of Mexico, and it grows widely in South America, Europe, Africa and [Asia. It ha](#page-6-8)s [been](#page-6-8) [reported tha](#page-6-9)t [the m](#page-6-9)ain traditional use was as an anti-diarrheal. Gastroenteritis, dysentery, stomach disorders, antibacterial colic pathogenic germs of the intestine were the ailments for which this plant was used as a treatment (Gutiérrez *et al.*, 2008). The leaves were used to prepare a decoction to treat cough (Heinrich *et al.*, 1998). In Mexico it is used to treat gastrointestinal and respiratory disturbances and also as an anti-inflammatory [medic](#page-6-10)ine (Rehab *et al.*, 2019). *Psidium guajava* leaves have shown [to possess some ethn](#page-6-11)omedical uses such as to treat diarrhea and stomach in the form of infusion or decoction (Pontikis, 1996), Diabetes melli[tus, hypertension \(O](#page-7-5)h *et al.*, 2005; Ojewole, 2005), Febrifuge, antispasmodic, rheumatism and several other treatments. The leaves were chewed to relieve toothache an[d applied on wo](#page-7-6)unds, [ulcers and r](#page-7-7)heumatic pain (He[inrich](#page-6-12) *et al.*, [199](#page-6-12)[8\). It](#page-7-7)

is used as an astringent, drying agent and a diuretic in Latin America, Central and West Africa and in Southeast Asia. The decoction of the leaves and bark has been used in India for treating diarrhea, dysentery, vomiting, sore throat and menstrual cycle regulation. Amazon tribes have been using the decoction for the treatment of bleeding gums, rinse off vaginal discharge and to tone up the vaginal walls after labour (Kamath *et al.*, 2008).

Ryu *et al.* (2012) has reported that compounds such as *γ* (*γ*)-sitosterol, Vitamin E and Squalene present in *P.guajava* [contri](#page-6-13)bute to the potent anticancer [activity by ex](#page-6-13)hibiting certain mechanisms s[uch as sup](#page-7-8)p[ressio](#page-7-8)n of signalling pathways, apoptosis induction and cell-cycle arrest. *β*-caryophyllene inhibits the lipopolysaccharide-stimulated proinflammatory cytokines (TNF- α and IL-1 β) in peripheral blood, thereby exhibiting an anti-inflammatory efficacy (Gertsch et al., 2008). The study conducted by Ujiki *et al.* (2006) has shown the role of Apigenin in the inhibition of pancreatic cancer by suppressing the DNA synthesis and proliferation, G2/M stage, cy[clin B associated cdc](#page-6-14)2 activity in these pancre[atic cancer](#page-7-9) c[ell line](#page-7-9)s. Gemcitabine has been used as a drug of choice for pancreatic cancer either alone or with a combination of drugs (Orlandi *et al.*, 2016). The compound quercetin is known to possess antioxidant capacity (Thaipong *et al.*, 2005), hypoglycemic and anti-hypotensive effects (Ojewole, 2005; Wang *et al.*, 2005) .

[Thus,](#page-7-4) pancreatic cancer ha[s been seen to](#page-7-10) h[ave an](#page-7-10) aggressive nature and needs more specific tar[geting](#page-7-7) [of the dru](#page-7-7)[g to bring about a](#page-7-11)n effective inhibition of cancer. This study focuses on the proteins expressed in pancreatic cancer as a target for the in silico molecular interaction with the active compounds present in *P. guajava* to find out the best binding compounds. The results of this study can be tested further through in vitro and in vivo approaches to bring out an efficient therapeutic molecule.

MATERIALS AND METHODS

Ligand preparation

The *P. guajava* derived bioactive compounds were selected for the docking studies. Apigenin (CID: 5280443), *β* caryophyllene (CID: 5281515), *γ*sitosterol (clianosterol) (CID: 94195), Glycolic acid (CID: 757), Ledol (CID: 92812), Quercetin (CID: 5280343), Squalene (CID: 638072), Vitamin E (CID: 14985), Gemcitabine (Commercial drug) (CID: 60750) were retrieved from the PubChem database.

The ligands were converted from sdf to pdb format using PyMol software. The ligand was prepared for

docking using the Auto dock tools 4.2 software. Its torsions were detected and modified and saved as a pdbqt file. This file was used for docking.

Target protein preparation

A literature survey was made for the proteins expressed in pancreatic cancer in conditions such as metastasis and drug resistance. The proteins which were expressed in the pancreatic cancer were chosen to be docked. The proteins *β*-catenin (PDB ID: 3sl9), Mesothelin (PDB ID: 4f3f), CD43 (PDB ID: 1kyj), SNAI1 (PDB ID: 3w5k) P-selectin (PDB ID: 1g1q), HSF1 (PDB ID: 5hdg), CD44 (PDB ID: 1poz) were retrieved from RCSB PDB database, sequence for Claudin-4 (Uniprot ID: 014493) and PDGF-D (UniProt ID: Q9GZP0) were retrieved from Uniprot database and modelled with Swiss auto model and their 3D Structures were generated.

All proteins were prepared using the Auto dock tools 4.2 software and the pre-attached ligands and water molecules were removed and polar hydrogen and Kollman charges were added. And the file was saved as a new pdb file for docking. The grid box was set to the required size depending on the region of binding pockets present on that particular protein. The molecule was then used for docking.

Active site preparation

The active sites of the prepared target proteins were predicted using the Castp server (Tian *et al.*, 2018). The pdb file was uploaded in the webserver, and the results were obtained in which the active site residues were mentioned. The regions with these active site residues were chosen [for setting up th](#page-7-12)e grid box dimension in the Auto dock tools software.

Molecular docking

Auto dock vina was used for molecular docking and calculating binding affinities (Trott and Olson, 2009). Auto dock Vina operates via the command line terminal. The target protein name and the ligand name were specified with pdbqt extension in the input parameter under receptor a[nd ligand for auto](#page-7-13) [dock](#page-7-13) vina configuration, and the sizes and centers x, y, z, were mentioned for the grid parameters. These configurations were different proteins. Two separate files were created as output, one as a text log file and the other a pdbqt file after running the docking.

Visualization of docked structures

Biovia Discovery studio visualizer was used to analyze the docked structures obtained from Auto dock Vina. The ligand interactions were made visible and labelled with the amino acid residues. The binding affinities of the ligands (*P. guajava* derived bioactive compounds) were compared with that of the control ligand (Gemcitabine).

RESULTS AND DISCUSSION

Molecular docking

All docking results were retrieved in a log file in text format and output file in pdb format split by the command-line tool. Auto dock results output were obtained as Binding energy (kcal/mol) (negative value) in a log file and structure file (pdbqt). The binding scores of different ligands with different proteins were compared with the standard drug (gemcitabine). The binding energy less than standard drug was considered to be effective ligands and can be finalized for further studies.

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine **Figure 1: Molecular interaction between** *β***-Catenin and** *P. guajava* **derived bioactive compounds**

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

Analysis of docked structures

The docking results were analyzed using the Dis-

	Beta-	Claudin-4	Mesoth	$CD43-$	SNAI1	PDGF-	$P-$	HSF1	CD44
	catenin			mucin		D	selectin		
	(PDB ID:	(Uniprot)	(PDB	(PDB)	(PDB)	(Unipro	(PDB	(PDB)	(PDB
	3sl9	ID:	ID:	ID:	ID:	ID:	ID:	ID:	ID:
		014493)	4f3f	1kyj)	3w5k	Q9GZP0	1g1q	5hdg)	1poz)
Apigenin	-7.2	-5.6	-7.3	-5.7	-8.1	-7.8	-6.9	-5.5	-6.1
Beta-	-5.9	-5.5	-6.1	-5.7	-6.3	-5.8	-5.9	-4.7	-5.3
caryophyllene									
Gamma	-5.9	-6.0	-7.0	-5.7	-6.5	-5.9	-6.0	-5.5	-4.7
sitosterol									
(Clianos-									
terol)									
Glycolic	-3.9	-3.2	-4.1	-3.2	-4.0	-3.3	-3.7	-3.2	-3.6
acid									
Ledol	-5.7	-4.8	-5.4	-6.0	-6.5	-6.0	-5.3	-4.7	-5.5
Quercetin	-7.7	-6.4	-7.9	-6.3	-9.6	-6.9	-7.2	-6.2	-6.4
Squalene	-5.3	-4.2	-4.4	-3.4	-5.6	-3.8	-4.3	-3.7	-4.1
Vitamin E	-5.6	-5.7	-5.4	-4.4	-6.5	-6.1	-6.2	-4.6	-4.5
Gemcitabine	-6.5	-5.6	-6.3	-4.9	-6.9	-5.1	-6.3	-4.9	-5.6

Table 1: Binding scores (kcal/ mol) for the molecular interaction of bioactive compounds from *P. guajava* **with pancreatic cancer expressed proteins**

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine **Figure 4: Molecular interaction between Claudin-4 and** *P. guajava* **derived bioactive compounds**

covery studio viewer and the binding amino acid residues were found (Figure 1, Figure 2, Figure 3, Figure 4, Figure 5, Figure 6, Figure 7, Figure 8 and Figure 9)

From the results obtained fro[m](#page-2-0) molecu[la](#page-2-1)r docki[ng](#page-3-0), it was [fo](#page-3-1)und th[at](#page-4-0) querce[tin](#page-4-1), a co[mp](#page-4-2)ound d[er](#page-4-3)ived from t[he](#page-5-0) ethanolic extract of *P. guajava* leaves, has shown to have the highest binding affinity for 8 proteins, namely *β*-catenin, Claudin-4, Mesothelin, CD43-mucin, SNAI1, P-selectin, CD44, HSF1. Api-

genin has shown to have the highest binding affinity for PDGF-D protein.

The results of the interaction and binding affinities have been given below in the order of Figure 1: *β*catenin, Figure 2: CD43, Figure 3: CD44, Figure 4: Claudin-4, Figure 5: HSF1, Figure 6: Mesothelin, Figure 7: PDGF-D, Figure 8: P-selectin, Figure 9: SNAI1 along with the ligands in the order of, a. Quer[ce](#page-2-0)tin, b. Apigenin, c. *[β](#page-2-1)*-caryophyllene[, d](#page-3-0). *γ*-sitosterol, [e](#page-3-1). Led[ol](#page-4-2), f. Squalen[e,](#page-4-0) g. [V](#page-4-3)itamin E, [h](#page-4-1). Glyco[lic](#page-5-0) acid, i.

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine **Figure 5: Molecular interaction between HSF-1 and** *P. guajava* **derived bioactive compounds**

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

Figure 6: Molecular interaction between Mesothelin and *P. guajava* **derived bioactive compounds**

Gemcitabine in each image.

The molecular docking was performed using auto dock vina and the results of the dockings were analyzed using the discovery studio visualizer. The necessity of protein-ligand docking is to find the best fit of ligand to the protein's three-dimensional structure. Nine receptors (proteins) and nine ligands (bioactive compounds from *P. guajava*) were chosen for docking. Among the nine ligands docked, quercetin was the ligand that showed the best docking score for eight proteins, namely *β*catenin, Claudin-4, Mesothelin, CD43-mucin, SNAI1, P-selectin, CD44, HSF1 and apigenin was the ligand that showed the best docking score for the protein PDGF-D. The docking scores were represented in terms of binding affinity denoted by kcal/mol.

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. y-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

Figure 7: Molecular interaction between PDGF-D and *P. guajava* **derived bioactive compounds**

a. Quercetin, b. Apigenin, c. B-caryophyllene, d. y-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

Figure 8: Molecular interaction between P-selectin and *P. guajava* **derived bioactive compounds**

The docking between the protein SNAI1 and quercetin gave the best score of -9.6 kcal/mol. Quercetin bound to the amino acid residues CYS262, ARG264, ASP339, ASP426, ARG465, ASN469, SER527 of SNAI 1 with 4 H-bonds. The second highest binding affinity was found to be with the docked protein mesothelin with a score of -7.9 kcal/mol with the amino acid residues GLY41, GLY42, LYS43, VAL93 with 3 H-bonds. Apigenin docks with PDGF-D with a binding score of -7.8 at the amino acid residues VAL186, PRO203, LEU205, ALA215, GLU216 with 1 H- bond. Quercetin binds to β -catenin with a binding affinity of -7.7 kcal/mol at the amino acid residues ILE153, PRO154, ARG190, GLN193, ARG274, LEU279 with 3 H-bonds.

a. Quercetin, b. Apigenin, c. β-caryophyllene, d. γ-sitosterol, e. Ledol, f. Squalene, g. Vitamin E, h. Glycolic acid, i. Gemcitabine

Figure 9: Molecular interaction between SNAI1 and *P. guajava* **derived bioactive compounds**

Quercetin binds with the protein P-selectin with a binding affinity of -7.2 kcal/mol at the amino acid residues GLN30, ASN31, LEU134, THR136, ILE137, CYS142, PRO151 with 5 H-bonds. The binding affinities of the proteins claudin-4 and CD44 with quercetin were found to be the same – 6.4 kcal/mol with amino acid residues LYS103, GLU105, ALA115 with 2 H-bonds and TYR42, TYR79, ALA98, SER112 with 2 H-bonds, respectively. Quercetin docks with the protein CD43 at the amino acid residues GLN7, GLY102, LYS103, ALA113, ALA115, MET118 at 4 H-bonds with a binding affinity of -6.3. Quercetin binds with the least docking sore of -6.2 to the protein HSF1 at the amino acid residues VAL26, ASP32, TRP37, SER38, PRO39, ARG106 with 3 H-bonds. Following the least score with the HSF1 protein.

The commercial drug gemcitabine is commonly used for many types of cancer chemotherapy and especially for pancreatic cancer. This compound was found to have an efficient binding score of -6.9 kcal/mol for the protein SNAI1 with the amino acid residues ASP339, ARG465, SER468, ASN469 with 2 H-bonds. Gemcitabine showed the least binding affinities for two proteins, namely CD43 and HSF1 at the amino acid residues ASP76, ASN142, GLN145, ASP146, ARG158 with 2 H-bonds and MET75, TYR76, HIS101, LYS116, ARG117 at 1 Hbond with scores -4.9 for both the proteins (Table 1).

The compound quercetin is a major flavonoid belonging to the class of flavonols. It is mostly found in foods like cauliflower, nuts, tea, apples, gua[va](#page-3-2)s and berries.

The anti-oxidant activity of the quercetin is responsible for its radical scavenging, lipid peroxidation inhibition and metal chelation mechanisms in

vitro (Rice-Evans *et al.*, 1996). The drug-likeness of quercetin was very good which can be understood by its molecular properties. It has 5 hydrogen bond donors (nOHNH) and 7 hydrogen bond acceptors (nON) with an octal water partial coefficient $(\log n)$ P) of 1.683, having a molecular weight of 302.238 g/mol, 22 non-hydrogen atoms, a single rotatable bond, the polar surface area of 131.351 A2 and a molecular volume of 240.084.

The overall drug-likeness score for quercetin was reported 1.00 which is actually a good score (Islam *et al.*, 2013). Compound quercetin was found to have a binding affinity of $-$ 4.52 kcal/mol against cellular tumor antigen p53 in a human cervical cancer cell line (HELA) and bound with NF- kappa [B with](#page-6-15) [the least sc](#page-6-15)ore of – 2.83 kcal/mol. (Muthukala *et al.*, 2015).

Furthermore, lung cancer proteins viz., p53, caspase 3 and mucosal addressin cell adh[esion molecule 1](#page-6-16) [were](#page-6-16) targeted for apigenin docking, which showed $-$ 4.6 kcal/mol of binding affinity against the p53 protein, - 5.7 kcal/mol against caspase3, and – 5.3 kcal/mol against mucosal addressin cell adhesion molecule 1 (Kasilingam and Elengoe, 2018) and a drug-likeness score of apigenin was found to be 0.940 (Hashemi, 2012).

From the a[bove information on the per](#page-6-17)formed molecular docking studies and the available literature, it [is understood t](#page-6-18)hat the compounds quercetin and apigenin have been found to be potent anticancer agent against pancreatic cancer and can be therapeutically effective.

CONCLUSION

The overall studies on the in silico molecular interaction between pancreatic cancer expressed proteins and *P. guajava* derived bioactive compounds using the Auto dock vina has been carried out efficiently and the binding affinities of the compounds have been identified. Quercetin and apigenin have been found to be the compounds with good binding affinities towards pancreatic cancer expressed proteins. Therefore, it has been well understood from the above approaches that specific compounds such as quercetin and apigenin present in the *P. guajava* have the potential to be developed as a very effective anti-cancer drug in future, provided its solubility, pharmacokinetics and toxicity level must be optimized through dry and wet lab approaches. Further studies will look forward to investigating this anticancer activity in depth by molecular dynamics simulation, in vitro and in vivo studies and gene expression analysis.

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Conϐlict of Interest

The authors declare that they have no conflict of interest for this study.

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REFERENCES

- Dorjee, P., Long, Z.-W. 2018. A mixed treatment comparison of toxicity of gemcitabine combined with different targeted drugs in the treatment of advanced or metastatic pancreatic cancer. *Cancer Biology & Therapy*, 19(6):497–506.
- Gertsch, J., Leonti, M., Raduner, S., Racz, I., Chen, J.- Z., Xie, X.-Q., Altmann, K.-H., Karsak, M., Zimmer, A. 2008. Beta-caryophyllene is a dietary cannabinoid. *Proceedings of the National Academy of Sciences*, 105(26):9099–9104.
- Gilabert, M., Chanez, B., Rho, Y. S., Giovanini, M., Turrini, O., Batist, G., Kavan, P., Raoul, J. L. 2017. Evaluation of gemcitabine efficacy after the FOLFIRI-NOX regimen in patients with advanced pancreatic adenocarcinoma. *Medicine*, 96(16):e6544–e6544.
- Gottesman, M. M., Fojo, T., Bates, S. E. 2002. Multidrug resistance in cancer: role of ATP– dependent transporters. *Nature Reviews Cancer*, 2(1):48–58.
- Gutiérrez, R. M. P., Mitchell, S., Solis, R. V. 2008. Psidium guajava: A review of its traditional uses, phytochemistry and pharmacology. *Journal of Ethnopharmacology*, 117(1):1–27.
- Hanahan, D., Weinberg, R. A. 2011. Hallmarks of Cancer: The Next Generation. *Cell*, 144(5):646–674.
- Hashemi, M. 2012. Protein tyrosine kinase (PTK) as a novel target for some natural anti-cancer molecules extracted from plants. *Journal of Medicinal Plants Research*, 6(27):4375–4378.
- Heinrich, M., Ankli, A., Frei, B., Weimann, C., Sticher, O. 1998. Medicinal plants in Mexico: healers' consensus and cultural importance. *Social Science & Medicine*, 47(11):1859–1871.
- Hung, S. W., Mody, H. R., Govindarajan, R. 2012.

Overcoming nucleoside analog chemoresistance of pancreatic cancer: A therapeutic challenge. *Cancer Letters*, 320(2):138–149.

- Islam, M. R., Zaman, A., Jahan, I., Chakravorty, R., Chakraborty, S. 2013. In silico QSAR analysis of quercetin reveals its potential as therapeutic drug for Alzheimer's disease. *Journal of Young Pharmacists*, 5(4):173–179.
- Jäger, W. 2009. Classical resistance mechanisms. *Int. Journal of Clinical Pharmacology and Therapeutics*, 47(01):46–48.
- Kamath, J. V., Rahul, N., Kumar, C. A., Lakshmi, S. M. 2008. Psidium Guajava L: A Review. *International Journal of Green Pharmacy*, 2(1):9–12.
- Kasilingam, T., Elengoe, A. 2018. In silico molecular modeling and docking of apigenin against the lung cancer cell proteins. *Asian Journal of Pharmaceutical and Clinical Research*, 11(9):246–252.
- Kasuya, K., Tsuchida, A., Nagakawa, Y., Suzuki, Y., Suzuki, M., Aoki, T., Abe, Y., Shimazu, M., Itoi, T., Sofuni, A. 2011. Prediction of a Side Effect and Efficacy of Adjuvant Chemotherapy with Gemcitabine for Post Operative Patient of Pancreatic Cancer by a Genetic Polymorphism Analysis. *Hepatogastroenterology*, 59(117):1609–1613.
- Kobayashi, N., Shimamura, T., Tokuhisa, M., Goto, A., Endo, I., Ichikawa, Y. 2017. Effect of FOLFIRINOX as second-line chemotherapy for metastatic pancreatic cancer after gemcitabine-based chemotherapy failure. *Medicine*, 96(19):e6769– e6769.
- Long, J., Zhang, Y., Li, M. 2011. Overcoming drug resistance in pancreatic cancer. *Expert Opinion on Therapeutic Targets*, 15(7):817–828.
- Muranaka, T., Kuwatani, M., Komatsu, Y., Sawada, K., Nakatsumi, H., Kawamoto, Y., Yuki, S., Kubota, Y., Kubo, K., Kawahata, S., Kawakubo, K., Kawakami, H., Sakamoto, N. 2017. Comparison of efficacy and toxicity of FOLFIRINOX and gemcitabine with nabpaclitaxel in unresectable pancreatic cancer. *Journal of Gastrointestinal Oncology*, 8(3):566–571.
- Muthukala, B., Sivakumari, K., Ashok, K. 2015. In Silico Docking of Quercetin Compound Against the Hela Cell Line Proteins. *International Journal of Current Pharmaceutical Research*, 7:13–16.
- Nakano, Y., Tanno, S., Koizumi, K., Nishikawa, T., Nakamura, K., Minoguchi, M., Izawa, T., Mizukami, Y., Okumura, T., Kohgo, Y. 2007. Gemcitabine chemoresistance and molecular markers associated with gemcitabine transport and metabolism in human pancreatic cancer cells. *British Journal of Cancer*, 96(3):457–463.
- Oh, W. K., Lee, C. H., Ahn, J. S. 2005. Antidiabetic

effects of extracts from Psidium guajava. *Journal of Ethnopharmacology*, 96(3):411–415.

- Ojewole, J. A. O. 2005. Hypoglycaemic and hypotensive effects of Psidium gajava Linn. (Myrtaceae) leaf aqueous extract. *Methods and Findings in Experimental and Clinical Pharmacology*, 27(10):689–689.
- Orlandi, A., Calegari, M. A., Martini, M., Cocomazzi, A., Bagalà, C., Indellicati, G., Zurlo, V., Basso, M., Cassano, A., Larocca, L. M., Barone, C. 2016. Gemcitabine versus FOLFIRINOX in patients with advanced pancreatic adenocarcinoma hENT1-positive: everything was not too bad back when everything seemed worse. *Clinical and Translational Oncology*, 18(10):988–995.
- Pontikis, C. A. 1996. Psidium guajava L. (Guava) En. *Biotechnology in agriculture and forestry*, 35(4).
- Rahib, L., Smith, B. D., Aizenberg, R., Rosenzweig, A. B., Fleshman, J. M., Matrisian, L. M. 2014. Projecting Cancer Incidence and Deaths to 2030: The Unexpected Burden of Thyroid, Liver, and Pancreas Cancers in the United States. *Cancer Research*, 74(11):2913–2921.
- Rehab, H. A., Mostafa, M., Heba, E. M. 2019. Green Synthesis of Silver Nanoparticles Using Psidumguajava leaf Extract. *Journal of Environmental Science*, 46(1):1–19.
- Rice-Evans, C. A., Miller, N. J., Paganga, G. 1996. Structure-antioxidant activity relationships of flavonoids and phenolic acids. Free Radical *Biology and Medicine*, 20(7):933–956.
- Ryu, N. H., Park, K. R., Ahn, K. S. 2012. A Hexane Fraction of Guava Leaves (Psidium guajava L.) Induces Anticancer Activity by Suppressing AKT/Mammalian Target of Rapamycin/Ribosomal p70 S6 Kinase in Human Prostate Cancer Cells. *Journal of Medicinal Food*, 15(3):231–241.
- Street, W. 2019. Cancer facts and figures. *American Cancer Society*, 76.
- Thaipong, K., Boonprakob, U., Cisneros-Zevallos, L., Byrne, D. H. 2005. Hydrophilic and lipophilic antioxidant activities of guava fruits. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 36(Supple 4):254–257.
- Tian, W., Chen, C., Lei, X., Zhao, J., Liang, J. 2018. CASTp 3.0: computed atlas of surface topography of proteins. *Nucleic Acids Research*, 46(W1):W363–W367.
- Trott, O., Olson, A. J. 2009. AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2):455–461.
- Ujiki, M. B., Ding, X. Z., Adrian, T. E. 2006. Apigenin inhibits pancreatic cancer cell proliferation through G2/M cell cycle arrest. *Molecular Cancer*, 5(1):76–76.
- Wang, B., Liu, H. C., Ju, C. Y. 2005. Study on the hypoglycemic activity of different extracts of wild Psidium guajava leaves in Panzhihua Area. *Medical Science Edition*, 36(6):858–861.
- Wang, Z., Li, Y., Sarkar, F. H. 2011. Pancreatic cancer: understanding and overcoming chemoresistance. *Nature Reviews Gastroenterology and Hepatology*, 8(1):27–33.
- Warshaw, A. L., del Castillo, C. F. 1992. Pancreatic Carcinoma. *New England Journal of Medicine*, 326(7):455–465.